



Prenatal and recent methylmercury exposure and heart rate variability in young adults: the Seychelles Child Development Study

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Abstract

Epidemiologic evidence of an adverse association between exposure to methylmercury (MeHg) from consuming fish and heart rate variability (HRV) is inconclusive. We aimed to evaluate MeHg exposure in relation to HRV parameters in a large cohort of young adults from a high fish consuming population in the Republic of Seychelles. Main Cohort participants in the Seychelles Child Development Study were evaluated at a mean age of 19 years. Prenatal MeHg exposure was determined in maternal hair growing during pregnancy and recent exposure in participant's hair taken at the evaluation. The evaluation consisted of short (~2 hours) and long (overnight) Holter recordings obtained in 514 and 203 participants, respectively. Multivariable analyses examined the association of prenatal and recent MeHg exposure (in separate models) with time-domain and frequency-domain HRV parameters in different physiologic circumstances: supine position, standing position, mental stress when undergoing a mathematics test, sleep, and long recording. Prenatal MeHg exposure was not associated with any of the 23 HRV parameters studied after adjustment for multiplicity. The recent MeHg showed a trend toward significance only for few variables in the primary model. However, after additional adjustment for activity levels, polyunsaturated fatty acids, and multiplicity none were significant after a Bonferroni adjustment. In conclusion, prenatal and recent MeHg exposure had no consistent pattern of associations to support the hypothesis that they are adversely associated with heart rate variability in this study population that consumes large amounts of fish.

Keywords	mercury, methylmercury, heart rate variability, autonomic nervous system, fish consumption
Taxonomy	Neuroscience, Toxicology
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Corresponding Author	Wojciech Zareba
Order of Authors	Wojciech Zareba, Sally W. Thurston, Grazyna Zareba, Jean-Philippe Couderc, Katie Evans, Xiaojuan Xia, Gene Watson, JJ Strain, Emeir McSorley, Alison Yeates, Maria Mulhern, Conrad Shamlaye, Pascal Bovet, EDWIN VAN WIJNGAARDEN, Philip Davidson, Gary Myers

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Research Data Related to this Submission

There are no linked research data sets for this submission. The following reason is given:
Data will be made available on request

Highlights

- High fish consumption might be associated with exposure to methylmercury.
- The effect of methylmercury on heart rate variability remains controversial.
- This is the most comprehensive assessment of HRV parameters to our knowledge among published studies focused on methylmercury toxicity
- In this study, methylmercury exposure from open ocean fish consumption did not adversely impact heart rate variability in young adults.

1 **Prenatal and Recent Methylmercury Exposure and Heart Rate Variability in Young Adults:**
2 **the Seychelles Child Development Study**

3

4 ***Short Title: Methylmercury and HRV in Young Adults***

5 Wojciech Zareba¹, Sally W. Thurston,² Grazyna Zareba,³ Jean Philippe Couderc,¹ Katie Evans,²
6 Jean Xia,¹ Gene E Watson,^{3, 10} JJ Strain,⁴ Emeir McSorley,⁴ Alison Yeates,⁴ Maria Mulhern,⁴
7 Conrad F Shamlaye,⁵ Pascal Bovet,⁶, Edwin van Wijngaarden,^{3,7,8, 10} Philip W Davidson,^{3,8} Gary
8 J Myers^{8,3,9}

9

10 **From:**

11 (1) Heart Research, Cardiology Division; University of Rochester Medical Center, Rochester,
12 NY

13 (2) Department of Biostatistics and Computational Biology; University of Rochester Medical
14 Center, Rochester, NY

15 (3) Department of Environmental Medicine, University of Rochester Medical Center, Rochester,
16 NY;

17 (4) University of Ulster, Coleraine, Northern Ireland, UK;

18 (5) Ministry of Health, Republic of Seychelles, Seychelles;

19 (6) University Institute of Social and Preventive Medicine, Lausanne, Switzerland

20 (7) Department of Public Health Sciences; University of Rochester Medical Center, Rochester,
21 NY

22 (8) Department of Pediatrics, University of Rochester Medical Center, Rochester, NY

23 (9) Department of Neurology, University of Rochester Medical Center, Rochester, NY

24 (10) Eastman Institute for Oral Health, University of Rochester, Rochester, NY

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30 Address for Correspondence:

31

32 Wojciech Zareba, MD, PhD

33 Cardiovascular Clinical Research Center, Cardiology Division

34 University of Rochester Medical Center

35 265 Crittenden Blvd.

36 Rochester, NY 14642

37 e-mail: wojciech_zareba@urmc.rochester.edu

39 **ABSTRACT**

40 Epidemiologic evidence of an adverse association between exposure to methylmercury (MeHg)
41 from consuming fish and heart rate variability (HRV) is inconclusive. We aimed to evaluate MeHg
42 exposure in relation to HRV parameters in a large cohort of young adults from a high fish
43 consuming population in the Republic of Seychelles. Main Cohort participants in the Seychelles
44 Child Development Study were evaluated at a mean age of 19 years. Prenatal MeHg exposure
45 was determined in maternal hair growing during pregnancy and recent exposure in participant's
46 hair taken at the evaluation. The evaluation consisted of short (~2 hours) and long (overnight)
47 Holter recordings obtained in 514 and 203 participants, respectively. Multivariable analyses
48 examined the association of prenatal and recent MeHg exposure (in separate models) with time-
49 domain and frequency-domain HRV parameters in different physiologic circumstances: supine
50 position, standing position, mental stress when undergoing a mathematics test, sleep, and long
51 recording. Prenatal MeHg exposure was not associated with any of the 23 HRV parameters
52 studied after adjustment for multiplicity. The recent MeHg showed a trend toward significance
53 only for few variables in the primary model. However, after additional adjustment for activity
54 levels, polyunsaturated fatty acids, and multiplicity none were significant after a Bonferroni
55 adjustment. In conclusion, prenatal and recent MeHg exposure had no consistent pattern of
56 associations to support the hypothesis that they are adversely associated with heart rate
57 variability in this study population that consumes large amounts of fish.

59 Fish is an important source of nutrition worldwide with about 3.1 billion people depending
60 daily on fish as their primary source of protein (FAO 2016). However, all fish also contain naturally
61 occurring methylmercury (MeHg) which is a known neurotoxicant in high dosages. Maternal
62 consumption during pregnancy of industrially polluted fish (after MeHg was discharged in waste
63 water from a chemical plant and seed grain treated with a MeHg fungicide) have been associated
64 with severe neurological deficits in children (WHO 2007). Since neurotoxicity was observed after
65 large exposure to MeHg, several studies have evaluated heart rate variability (HRV) to investigate
66 the association between prenatal or recent MeHg exposure (Sorensen et al. 1999; Oka et al.
67 2002; Grandjean et al. 2004; Murata et al. 2004; Choi et al. 2009; Lim, Chung, and Paek 2010).

68 Heart rate variability (HRV) is an accepted method to assess changes in the autonomic
69 nervous system during parasympathetic and sympathetic modulation of the cardiovascular
70 system (HRV 1996). Heart rate is under constant influence of an interplay between
71 parasympathetic and sympathetic modulation leading to slower heart rate at rest or during sleep,
72 and faster heart rate with exercise or stress. Beat-to-beat variation in heart rate, or in duration of
73 beat-to-beat intervals, could be measured reflecting changes in the autonomic control of the heart
74 (Kleiger, Stein, and Bigger 2005). In healthy subjects, it is expected that there is a physiologic
75 variation of on NN intervals (defined as normal beats with sinus rhythm, while excluding premature
76 and ectopic beats) influenced by respiration, baroreflex sensitivity, in addition to the above
77 mentioned parasympathetic and sympathetic modulation (Cygankiewicz and Zareba 2013). HRV
78 might be altered in patients after myocardial infarction, patients with heart failure, or those with
79 diabetes, obesity, frequently with a decrease in parasympathetic modulation and an increased
80 sympathetic modulation (HRV 1996; El Agaty, Kirmani, and Labban 2017; Tsuji et al. 1996).
81 These changes in HRV have been found as predictors of outcome (mortality, heart failure,
82 ventricular tachyarrhythmias) in various patient populations (Kleiger et al. 1987; Bigger et al. 1992;

83 Stein et al. 1997; Kleiger, Stein, and Bigger 2005; Sherazi et al. 2015). There is profound evidence
84 that in addition to environmental factors like exposure to air pollution or lead, pharmacological or
85 electrical therapies might alter HRV parameters indicating changes in the autonomic control of
86 the heart (Coumel et al. 1991; Malik et al. 1989; Simula et al. 2017; Jerling et al. 2018;
87 Hoogerwaard et al. 2019; Petersen et al. 2018; Breitner et al. 2019; Gump et al. 2011). The
88 guidelines regarding HRV analysis and its parameters recommended for research and clinical
89 use were established in 1996 (HRV 1996). However, over the last 20 years there still have been
90 inconsistencies regarding HRV methodology and interpretation (Shaffer and Ginsberg 2017).

91 Some epidemiologic studies have suggested that MeHg exposure may be associated with
92 a decrease in HRV parameters but with general inconsistency of findings among studies,
93 (Grandjean et al. 2004; Mozaffarian 2009; Oka et al. 2002; Sorensen et al. 1999) and some were
94 not able to confirm it (Valera, Dewailly, and Poirier 2008; Miller et al. 2018; Gump et al. 2017;
95 Choi et al. 2009) These studies have generally been based on short term HRV recordings at rest,
96 and some were done in populations with co-exposures. Varying duration of recordings, diverse
97 ECG sampling rate, different management of ectopic beats may influence results of HRV analyses
98 (Shaffer and Ginsberg 2017; Karita et al. 2018; Stapelberg et al. 2018; Melo et al. 2018). These
99 factors limit our understanding of whether there are systematic and direct cardiovascular effects
100 of MeHg exposure (Karita et al. 2018). As recently summarized in two extensive reviews on the
101 topic (Gribble et al. 2015; Karita et al. 2018) it is still uncertain whether MeHg exposure from
102 repeated consumption of fish with naturally occurring background levels has any adverse and
103 meaningful effect on HRV.

104 In a prospective study of 19-year-old healthy subjects, we assessed whether the prenatal
105 and recent MeHg exposure from consuming fish adversely affects HRV in comprehensive
106 analyses of 24-hour Holter recordings and short-term 5-minute recordings reflecting different

107 physiologic conditions associated with resting supine, standing, mental stress (mathematics test),
108 and sleeping.

109

110 **METHODS**

111 **Study Population and Design**

112 The Republic of Seychelles is a 115-island archipelago in the middle of the Indian Ocean
113 where citizens consume large amounts of ocean fish. The population is exposed to MeHg
114 primarily from fish consumption and does not consume sea mammals (Cernichiari et al. 1995;
115 Davidson et al. 1998). Fish consumed in Seychelles are contaminated only by natural
116 background exposure to MeHg. This setting provides a unique opportunity to study the
117 association of MeHg with health outcomes. In 1989-90, 779 mother-infant pairs were recruited
118 as part of the Seychelles Child Development Study (SCDS) main cohort (Marsh et al. 1995). The
119 demographics and earlier evaluations of this cohort have been reported previously (Davidson et
120 al. 1998; Shamlaye et al. 1995; Myers et al. 2003; van Wijngaarden et al. 2013).

121 For this study, a total of 518 cohort participants returned for evaluations at the Study
122 Centre at a mean \pm SD of 19.0 ± 0.3 years of age. Data from four of these participants were not
123 included due to technical errors in ECG data acquisition. The evaluation consisted of a
124 questionnaires collecting demographic, socioeconomic, medical history, and lifestyle data,
125 collection of biological samples (hair and blood samples), and Holter ECG recordings providing
126 time- and frequency-domain HRV parameters. The study protocol was approved by the
127 Institutional Review Boards of the Republic of Seychelles and the University of Rochester.

128 **MeHg and Polyunsaturated Fatty Acids Measurements**

129 Prenatal MeHg exposure was determined by measurement of total maternal Hg levels in
130 hair in samples collected close to the scalp after pregnancy. It reflected mercury concentrations
131 during pregnancy assuming a hair growth rate of 1 cm per month. Recent MeHg exposure was
132 available for 451 (87.7%) of the participants; 68% of those missing this variable were males, many
133 of whom had shaved heads. Recent MeHg was analyzed in the 1 cm hair samples collected
134 closest to the scalp taken at the time of Holter testing (Myers et al. 2009). Hair analysis was
135 performed using cold vapor atomic absorption spectroscopy (Magos and Clarkson 1972). The
136 detailed description of hair sampling, analysis and quality control procedures was previously
137 described (Cernichiari et al. 1995). It has been documented that mercury hair levels recapitulate
138 mercury exposure since hair concentrations correlate well with blood mercury levels (Clarkson,
139 Magos, and Myers 2003).

140 Recent status of n-3 and n-6 PUFA were determined from plasma samples taken at the
141 time of evaluation. Samples were processed, aliquoted, and stored at -80°C in the Ministry of
142 Health's Clinical Laboratories and transported to the nutrition biochemistry laboratory in
143 Coleraine, Northern Ireland for fatty acid analysis. Plasma samples were subjected to lipid
144 extraction using an adaptation of the method of Folch et al. (Folch, Lees, and Sloane Stanley
145 1957) and solid phase extraction columns (Merck Lichrolut, UK) were used to isolate
146 phospholipids. Fatty acid methyl esters were prepared with boron trifluoride in methanol (Sigma-
147 Aldrich Co. Ltd.) and analyzed using GC-MS technology (5975C series, Agilent Technologies
148 Ltd., Stockport, UK). Fatty acids were identified by their retention time with reference to
149 commercially obtained fatty acid standards and were quantified by use of an internal standard
150 C17:0 (Sigma Aldrich Co. Ltd.). The GC column used was a BPX 70 capillary column
151 (Phenomenex, Cheshire, UK) with a length of 100 m, internal diameter of 0.25 mm and a capillary
152 thickness of 0.25 µm.

153 **Holter ECG Recordings**

154 All 514 participants underwent 12-lead high-resolution Holter ECG recordings which were
155 initiated between 8-10 am using digital (1000 Hz) recorders (H-Scribe, Mortara Instruments Inc.,
156 Milwaukee, WI). ECG recordings started with a 10-minute supine position followed by a 10-minute
157 standing and subsequently a 10-minute ECG while the participant was completing mathematics
158 (math) test. The supine recording served to analyze HRV parameters in the resting non-active
159 state (parasympathetic dominance), the standing recording provided insight into the response of
160 the heart rate and HRV parameters to activation of the autonomic system by changing body
161 position (sympathetic activation), and the recording during a mathematical test reflected the effect
162 of mental stress on the studied ECG parameters (Tsuji et al. 1996; Umetani et al. 1998). The
163 Holter recordings were finished after 1-2 hours in 312 subjects, but in the remaining 202 subjects
164 recordings were continued to obtain long-term overnight data to further characterize the normal
165 physiologic predominance of parasympathetic modulation of the autonomic nervous system
166 during sleep and provide insight into the overall long-term values of time-domain HRV
167 parameters.

168 All recordings were sent to and analyzed at the University of Rochester Medical Center's
169 ECG Core Laboratory by technicians blinded to participants' demographic and methylmercury
170 exposure data. Routine beat annotation was performed identifying normal sinus beats and
171 abnormal beats (atrial and ventricular ectopic beats, noise) and only normal sinus beats were
172 included in the analyses. In the case of ectopic beats, the sinus normal-to normal beat (NN)
173 interval following the ectopic beat was not included. All recordings were verified by a cardiologist
174 (WZ).

175

176 **Heart Rate Variability**

177 From short-term recordings obtained in supine and standing positions and during the math
178 test, 5-minute ECG recordings were extracted starting 3 minutes from the beginning of each
179 segment to allow for adaptation to the changed conditions. A 5-minute segment from the
180 overnight Holter recording was also analyzed at approximately 2 am to represent HRV during
181 sleep.

182 Annotated NN intervals were used to compute HRV parameters for a given segment of
183 data. The time-domain HRV parameters (Kleiger, Stein, and Bigger 2005) that were obtained from
184 both the short (~5-minute) and long recordings included: SDNN (standard deviation of normal-to-
185 normal sinus beat intervals) reflecting overall HRV and rMSSD intervals (root mean square of
186 successive differences in NN intervals). We also computed: pNN50 (percentage of NN intervals
187 differing more than 50 ms from the preceding beat) reflecting parasympathetic modulation,
188 SDANN (standard deviation of averaged 5-min normal-to-normal RR intervals), and SDNIX (mean
189 of the SDs of all normal-to-normal RR intervals) for all 5-min segments of the entire recording
190 (SDNNIX). These reflect overall HRV and are similar to SDNN, which is why we elected to focus
191 on SDNN and rMSSD as representative HRV parameters in time-domain analysis.

192 The frequency-domain HRV methods (Bigger et al. 1992) allow for identifying the relative
193 contribution of specific frequency bands reflecting oscillatory behavior of the heart rate. The total
194 power (TP) of the entire spectrum was measured from 0 to 0.40 Hz. Other components included
195 high frequency (HF: 0.15-0.40 Hz), low frequency (LF: 0.04-0.15 Hz), and very low frequency
196 (VLF: 0.003-0.04 Hz) bands. The HF power (expressed in normalized units as HF/[TP-VLF])
197 represents parasympathetic (respiratory) modulation of the heart rate whereas LF power
198 (expressed in normalized units as LF/[TP-VLF]) reflects modulation of the heart rate by both
199 sympathetic and parasympathetic tones, but with strong dominance of sympathetic influence and

200 baroreflexes. We also evaluated LF/HF ratio but elected to focus on LF and HF separately to
201 illustrate effects of MeHg on individual components of these variables.

202 Our primary statistical analysis focused on 23 of the most commonly used HRV
203 parameters reflecting autonomic activity in different physiologic conditions: 5 in supine position, 5
204 in standing position, 5 in response to a mathematics test, 5 during sleep, and 3 for the entire
205 recording. Five outcomes measured at baseline while supine at rest included: NN, SDNN, rMSSD,
206 LF, and HF. We also analyzed the five outcomes measured at baseline minus their values while
207 standing, taking a math test, and sleeping. Unless otherwise noted, response variables are
208 defined as differences from baseline values. Three primary outcomes measured on the long
209 recordings included: NN, SDNN, rMSSD. Additionally, we computed three more HRV parameters
210 (pNN50, SDANN, and SDNNIX) for secondary analysis by MeHg categories.

211

212 **Statistical Analyses**

213 We used linear regression to examine the association of each exposure (prenatal and
214 recent MeHg, in separate models) and each of the 23 HR variables. Models first examined the
215 association with a sex by exposure interaction term since significant differences in HRV are known
216 to be present between males and females, and additionally, exposure differed by sex (Tsuji et al.
217 1996; Umetani et al. 1998). If the interaction was statistically significant ($p < 0.05$) then those
218 results are reported. If the interaction was not significant ($p \geq 0.05$) then the analysis was rerun
219 without the interaction term and those results are reported. Primary models were fit with and
220 without covariate adjustment for sex, birth weight, and current BMI. We adjusted for birth weight
221 due to its potential impact on childhood development and for BMI due to evidence of an
222 association between ECG variables and obesity (Antelmi et al. 2004). For each analysis, we used

223 a two-sided test with $\alpha = 0.05$ to assess significance. For the association of recent MeHg
224 exposure with HRV we fit secondary models adjusting for recent n-3 and n-6 PUFA and for the
225 participant's activity level. Physical activity level is a main predictor of HRV (Fisher, Young, and
226 Fadel 2015). The Global Physical Activity Questionnaire (GPAQ) was developed by WHO for
227 physical activity surveillance. The GPAQ standardizes level of total physical activity measured in
228 MET-minutes/week categorizing the value <600 as low sedentary activity, $600-<3000$ as
229 moderate activity, and ≥ 3000 as high activity. The activity measurement was based on the
230 participants self-report of their minutes of physical activity per week using three domains (work,
231 walking, and leisure) and was treated as a categorical variable with three levels (<600 , $600-<3,000$,
232 $> 3,000$ minutes) following the WHO recommendation. We adjusted for n-3 and n-6
233 PUFA because of their reported cardiac effects (Christensen et al. 1999). Model assumptions
234 were checked using standard methods, including checking for constant variance, nonlinearity,
235 and normally distributed residuals (Weisberg 2005). In some cases, there was evidence of
236 assumption violations which necessitated a log transformation of the outcome. For variables that
237 required a logarithmic transformation, differences from baseline values were calculated after the
238 log transform of both variables (e.g. the outcome is the logarithm of the ratio of values, rather than
239 the logarithm of the difference from baseline). After transformations when required, there was no
240 evidence of extreme outliers or highly influential observations. We report the individual
241 associations and corresponding betas and p values from the primary analysis. Additionally, we
242 applied the Bonferroni correction to account for multiple comparisons. Since we focused on 23
243 primary HRV parameters measured (5 in supine position, 5 in standing position, 5 in response to
244 math test, 5 during sleep and 3 for the entire recording), we considered a p value of $0.05/23 =$
245 0.0022 as significant after this correction.

246 We also performed a univariate comparison of studied HRV parameters by MeHg levels
247 categorized as 0 to <5 ppm, 5 to <10 ppm, and ≥10 ppm in prenatal and recent exposure. Median
248 value of MeHg level in hair of US females 16-49 years of age in the NHANES 1999-2000 was
249 0.19 µg/g with 95th percentile of 1.73 µg/g, reaching 2.75 µg/g in individuals eating fish ≥3 times
250 in the past 30 days (McDowell et al. 2004). Prenatal assessments of women from the Seychelles
251 high fish consumption population reported mean ± SD total hair MeHg of 6.85 ± 4.5 ppm and a
252 median value of 5.94 ppm (Myers et al. 2003). Therefore, the cutoff of 0-5ppm was chosen as
253 reflecting low levels of MeHg. The median total hair Hg level of women in the Faroes birth cohort
254 study was 4.5 µg/g; 12% had levels > 10 µg/g, considered as elevated (Grandjean et al. 2004).
255 Therefore, we proposed the above arbitrary cutoffs reflecting low, medium, and high levels of
256 MeHg in the Seychelles fish eating populations.

257

258 **RESULTS**

259 **Descriptive Analysis: Characteristics of Participants**

260 There were 514 eligible participants (273 females and 241 males) who had short Holter
261 recordings and 203 participants (117 women and 86 men) who had long Holter recordings. Table
262 1 shows the participants clinical characteristics, MeHg levels, and n-3 and n-6 PUFA levels
263 compared by sex. The mean prenatal MeHg exposures were similar in males and females (6.77
264 vs 7.06 ppm, respectively), but recent MeHg exposures at 19 years of age were significantly
265 higher in males than females (12.16 vs. 8.67 ppm, respectively). Males were physically more
266 active and had significantly lower BMI values than females. The n-3 and n-6 PUFA levels did not
267 differ by sex. Correlation between prenatal Hg and recent Hg (obtained at the age of 19 years)
268 was weak: the correlation coefficient was 0.13.

269 As expected, time-domain SDNN and rMSSD parameters from long-term recordings and
270 short-term supine recordings were significantly ($p < 0.0022$) different between males and females
271 (Supplementary Table S1). In supine resting position, males had significantly longer NN interval
272 (lower heart rate) than females. Males also had significantly higher levels of long-term and
273 baseline supine time-domain SDNN and rMSSD parameters, but response to standing, math test,
274 and sleep did not significantly differ by sex. Differences in frequency-domain HRV parameters
275 between males and females were less pronounced and did not reach $p < 0.0022$ level.

276 **Heart Rate Variability Parameters by Categorized MeHg Exposure Levels**

277 Table 2 shows a univariate comparison of studied HRV parameters by MeHg levels
278 categorized as 0 to < 5 ppm (low), 5 to < 10 ppm (medium), and ≥ 10 ppm (high) in prenatal and
279 recent exposure (Table 2). For prenatal MeHg exposure this comparison did not show any
280 significant associations indicating level-related changes in HRV parameters in long term
281 recordings, baseline supine or response to standing, math and sleep. For recent MeHg categories
282 there were few statistically borderline associations (p value between 0.0022 and 0.05) which were
283 driven by lower values in medium than low and high MeHg exposure: NN in response to standing
284 and math test, LF in response to sleep, and HF at baseline, in response to math and sleeps.
285 Among these associations the direction was inconsistent and there was no evidence of a dose
286 response. Since we observed sex-related differences in HRV parameters we reanalyzed 24-hour
287 HRV data for males and females separately using the primary time domain parameters (NN,
288 SDNN, rMSSD) and 3 additional ones (pNN50, SDANN, & SDNNIX). No significant associations
289 were present in these HRV parameters among subject with low, medium, and high MeHg
290 exposure (Supplementary Table S2).

291 **Regression Analysis: Prenatal MeHg Exposure and Heart Rate Variability**

292 The sex by prenatal exposure interaction was significant for the NN interval long recording
293 and the NN in response to sleep (Table 3). After adjusting for sex, birth weight, and BMI, there
294 were two borderline significant associations at p value between 0.0022 and 0.05. Both were in
295 males; and one was beneficial and the other adverse. The NN interval in the long recording was
296 predicted to increase in males by 5.5 ms per 1 ppm increase in MeHg ($p=0.006$). This beneficial
297 association reflected a 0.5 beat per minute decrease in heart rate per 1 ppm increase in MeHg,
298 or a 3.4 beat per minute decrease per IQR increase in MeHg. The NN interval in response to
299 sleep decreased only in males by 10.49 ms per 1 ppm increase in MeHg ($p = 0.0055$), reflecting
300 about 0.5 a beat per minute increase in heart rate, physiologically an adverse association. When
301 the Bonferroni correction was applied, none of the associations reached statistical significance
302 ($p<0.0022$). There were no other significant interactions or associations of prenatal MeHg with
303 either the time- or frequency-domain HRV parameters computed from the long or short
304 recordings.

305 **Regression Analysis: Recent MeHg Exposure at 19 years and Heart Rate Variability**

306 Recent MeHg exposure was associated with four outcomes in models that adjusted for
307 sex, BMI, and birth weight (Supplementary Table S3). Only one sex by MeHg interaction was
308 borderline significant at p between 0.0022 and 0.05, the NN interval in response to standing ($p =$
309 0.015). That association was only in females and was adverse ($p = 0.0172$). For the NN interval
310 in response to standing, females had a 3 ms increase in the difference in the NN interval per 1
311 ppm increase of MeHg ($p = 0.017$), or a 20.8 ms increase in the difference in the NN interval for
312 an interquartile range (IQR). The associations with the two most classical time-domain
313 parameters: SDNN baseline supine and on the long recording were not significant ($p=0.600$ and
314 0.128 respectively). Similarly, the associations with rMSSD resting supine and on the long
315 recording were not significant ($p=0.197$ and 0.489 , respectively). For the difference in log

316 (rMSSD) in response to a math test, there was a 0.009 ms increase for each 1 ppm increase in
317 MeHg, a beneficial response. Stated differently, a 1 ppm increase in exposure was associated
318 with a 1% increase in the ratio of rMSSD at baseline to the rMSSD during the math test, or an
319 IQR increase in exposure was associated with a 6.4% increase in this ratio. For HF in response
320 to standing both sexes together had a 0.40% increase in difference from baseline ($p=0.029$), and
321 for HF in response to the math test, they had a 0.375% increase in difference from baseline
322 ($p=0.032$), indicating lower parasympathetic parameters during these tests. These associations
323 were inconsistent in direction, of small magnitude, nonsignificant after Bonferroni correction, and
324 of uncertain meaning.

325 After additional adjustment for activity levels and n-3 and n-6 PUFA, recent MeHg
326 exposure had borderline significant sex by recent MeHg interactions in two models: the NN
327 interval in response to standing and the LF in response to the math test (Table 4). The association
328 between MeHg and NN interval in response to standing was borderline at $p=0.0499$ only in
329 females who had on average a 2.6 ms increase per 1 ppm increase in recent MeHg. There was
330 also a borderline ($p=0.0210$) association between MeHg and LF in response to the math test for
331 males only. Each 1 ppm increase in exposure was associated with a 0.56 ms decrease in the
332 difference in LF (a beneficial effect).

333 Of the six significant borderline significant associations (at P between 0.0022 and 0.05)
334 associations with the combined sexes, three were also present in the primary analysis (the
335 difference in rMSSD on the logarithmic scale in response to the math test and the difference in
336 HF in response to standing and to the math test). The association with the difference in rMSSD
337 in response to the math test was virtually unchanged, and the associations with HF responses to
338 standing and the math test were both slightly attenuated. The three additional outcomes with
339 borderline significant MeHg associations were the NN interval for the long recording, the baseline

340 supine NN, and the LF in response to sleep. For the baseline supine NN, each 1 ppm increase
341 in recent MeHg was associated with on average a decrease of 3.05 ms, an adverse association.
342 For the LF in response to sleep, each 1 ppm increase in recent MeHg was associated with a 0.69
343 decrease (beneficial effect). The inconsistency of these associations and their small magnitude
344 (milliseconds) make them challenging to interpret. After applying the Bonferroni correction for
345 multiplicity, there were no statistically significant MeHg associations (all p values >0.0022).

346

347 **DISCUSSION**

348 We studied whether HRV parameters are associated with prenatal or recent MeHg
349 exposure. We measured a comprehensive set of HRV parameters obtained from short-term (5
350 minutes) and long-term (24 hours) Holter recordings in a cohort of young adults exposed to MeHg
351 from frequent consumption of open ocean fish. Their prenatal MeHg exposures were
352 approximately 10 times more than those present in the United States. We focused on 23
353 measured HRV variables and found none of them significant at prespecified after Bonferroni
354 correction with $p < 0.0022$. There were two borderline significant (at p between 0.0022 and 0.05)
355 associations with prenatal and eight borderline significant associations with recent MeHg
356 exposure in secondary models. The two prenatal associations were present only in males and
357 were in opposite directions suggesting random variation. The eight recent MeHg exposure
358 significant associations were all of small magnitude, and there was no consistent pattern that
359 would suggest they are related to exposure. Following Bonferroni correction for multiplicity, there
360 were no statistically significant (at $p < 0.0022$) associations with either prenatal or recent MeHg
361 exposure.

362 We observed the expected sex differences in HRV parameters that have been previously
363 reported in the literature. Males showed significantly higher values of HRV parameters than
364 females (Agelink et al. 2001; Umetani et al. 1998), and the responses to standing and other
365 conditions were as expected (Srinivasan, Sucharita, and Vaz 2002). These findings in a large
366 cohort provide reassurance that there was sufficient statistical power to detect meaningful
367 associations of exposures and HRV if they were present. The borderline significant findings were
368 of very small magnitude without clear physiological or clinical meaning. Previously, we measured
369 beat to beat blood pressure in 95 participants from this cohort, using the Finapres to record the
370 baroreflex, another measure of autonomic nervous system integrity (Periard et al. 2015). That
371 study found no association of blood pressure variability with either prenatal or recent MeHg levels.
372 These data do not support the hypothesis that prenatal or recent exposure to MeHg from fish
373 consumption influences HRV parameters.

374 The categorical analyses (Table 2 and Supplemental Table S2), although not adjusted for
375 covariates, further corroborate the absence of associations between prenatal and recent MeHg
376 exposure and the NN interval and other HRV parameters among the three exposure groups. If
377 there was an association, one would expect some MeHg level-dependent changes in HRV
378 parameters, but this categorical approach showed no trend toward any association, i.e. no
379 decrease or increase in studied parameters across incremental levels of MeHg. At the same time,
380 we were able to detect significant differences as we documented by detecting expected sex-
381 related differences in HRV parameters (positive control test).

382 Prenatal MeHg exposure was associated with about a 0.5 beat per minute slower heart
383 rate in males per 1 ppm increase in MeHg. This change does not have clinical meaning
384 considering that exposure to MeHg in Western societies is generally less than 1 ppm. Even in
385 high fish-consuming societies like Japan, MeHg exposure is generally less than 2 ppm which

386 would be associated with a potential variation in heart rate of approximately 1 beat per minute.
387 Neither time-domain nor frequency-domain HRV parameters were associated with prenatal MeHg
388 levels. Prenatal MeHg does not appear to be influencing parameters or autonomic nervous
389 system activity reflected by HRV at the age of 19 years in this cohort.

390 Several HRV parameters showed borderline significant associations after adjustment for
391 birth weight, BMI, activity level, and n-3 and n-6 PUFA levels. These associations were all of small
392 magnitude, inconsistent across parameters, and there was no indication of a dose response
393 curve. No associations were significant after Bonferroni adjustment. Additionally, it seems most
394 likely that if recent exposure to low levels of MeHg does impact the nervous system, it would be
395 associated with prolonged cumulative dosage rather than the one month value that our measure
396 determined. Considering all of these factors, we do not believe these data provide support for
397 the hypothesis that HRV parameters are associated with recent MeHg exposure.

398 In analyses adjusted for PUFA and activity level, heart rate assessed during supine,
399 standing, and math test periods increased with increasing MeHg levels with borderline
400 significance. A 3-millisecond decrease in NN interval translates to about 0.2- 0.3 beats per minute
401 increase in heart rate, and has no clinical relevance. Similarly rMSSD and HF decreased with
402 increasing recent MeHg during the math test and LF/HF with standing. These changes were very
403 small and not significant clinically or statistically after Bonferroni correction. In response to a math
404 test, there was a 0.010 ms decrease in rMSSD per 1 ppm increase in MeHg, which despite
405 borderline $p=0.006$ (after Bonferroni correction) is of no clinical or physiologic meaning.

406 Roman and colleagues reviewed the literature on associations of prenatal and recent
407 MeHg with cardiovascular outcomes (Roman et al. 2011). They noted that evidence of impact at
408 dose levels of interest was lacking and that the "...specific measures of HRV in each study vary."

409 However, based on the consistency of the epidemiological literature, they classified the evidence
410 supporting the association of HRV with MeHg exposure as “strong” compared to other
411 cardiovascular measures. This reasoning is tenuous when one considers the known bias against
412 publishing negative studies. The European Food Safety Authority ((CONTAM) 2012) reviewed
413 reports examining MeHg and HRV in 2012 with subsequent update in 2018 and the panel
414 concluded that, although some studies suggest an autonomic effect of MeHg, the results were
415 inconsistent among studies and the implication for health were unclear.

416 Recently, Japanese researchers (Karita et al. 2018) reviewed 13 studies examining the
417 association between MeHg and HRV parameters and 8 of them showed the significant
418 association with MeHg and 5 failed to demonstrate any significant association. Earlier studies
419 that reported an association between prenatal MeHg exposure and HRV parameters varied from
420 the current study in a number of ways. They were conducted in a variety of populations and
421 measured HRV variables using different methods and recording times. Also, some populations
422 studied had significant exposures to other toxicants that might also influence HRV.

423 Discussing studies supporting the MeHg and HRV associations, the Faroe Islands
424 investigators reported a decrease in the coefficient of variation of RR (a HRV parameter similar
425 to SDNN) and in LF and HF levels as prenatal MeHg increased (Sorensen et al. 1999; Grandjean
426 et al. 2004). Their studies differed in that Hg exposure was primarily from whale consumption
427 and were thus co-exposed to polychlorinated biphenyls. In addition, the ECG recordings were
428 short. The largest cohort studied to date evaluated 1,589 Koreans living near an industrial
429 complex. Participants ranged in age from 5 to 83 years with 389 below age 20 years, had mean
430 hair Hg levels that were low (0.83 ug/gm), and had 5-minute ECG recordings obtained in sitting
431 position, which affect results (Lim, Chung, and Paek 2010). When categorized by decades of life,
432 the 104 subjects below age 10 years had an 8.4% decrease in the HF parameter with each 1 ppm

433 increase in hair mercury concentration (95% confidence interval: 2.2-15.1). In a study of French
434 Polynesians aged 12–17 years (Valera et al. 2011), significant differences were observed in
435 LF/HF ratio, LF, and HF between the second (7.9–10.0 g/L) and third (11.0–26.0 g/L) tertiles of
436 blood mercury concentration. That study reported no associations between HRV parameters and
437 MeHg exposures in the 180 adults studied. In the only intervention study (Yaginuma-Sakurai et
438 al. 2010), a significant difference in LF was observed between the experimental group (mean
439 mercury levels of 8.76 µg/g in hair and 26.9 µg/L in blood) and control group, but one cannot
440 exclude the effect of concomitant changes in levels of n-3 PUFA influenced these results.

441 Discussing studies with no clear evidence of the MeHg-HRV association, analysis of
442 Faroese whaling men showed that blood mercury level was associated with increased coefficient
443 of variation of RR intervals, and coefficient of variation of LF, but latent mercury level, estimated
444 from mercury levels in blood, toe nail, and hair was not significantly associated with any HRV
445 parameter (Choi et al. 2009). In Inuit adults (Valera, Dewailly, and Poirier 2008), blood mercury
446 was significantly correlated with coefficient of variation of RR interval and LF in univariate
447 analyses, but these associations were not significant after adjusting for confounders. In fish
448 consumers from Long Island (Miller et al. 2018), HRV parameter in a multiple regression analysis
449 were not significant after adjustment for serum docosahexaenoic acid and eicosapentaenoic acid
450 levels. In the study of 203 children aged 9-11 years (Gump et al. 2017), LF, HF, and LF/HF ratio
451 at rest and during stress were not significantly associated with blood mercury levels.

452 As summarized above the effect of MeHg on HRV parameters does not seem consistent
453 and remains uncertain. Some studies indicate changes in the sympathetic (LF) component, some
454 in parasympathetic (HF, rMSSD), and some in both. Recording time and conditions varied among
455 the studies as did MeHg assessments. Competing environmental pollutions such as persistent
456 organic pollutants from consuming ocean mammals (Sorensen et al. 1999; Valera et al. 2011),

457 and underlying comorbidities such as in the Korean study (Lim, Chung, and Paek 2010), might
458 also have influenced the findings. If there is a causal association of prenatal or recent MeHg
459 exposure with changes in the autonomic nervous system, one would expect consistency in the
460 directionality of the associations for both classical time-domain and frequency-domain HRV
461 parameters. No study has demonstrated this. Our study with a large cohort and a wide range of
462 prenatal and recent MeHg exposures does not confirm the earlier reports.

463 We want to stress that none of the studies published to date in the literature on MeHg
464 association with HRV parameters has used 24-hour recording, the most standard approach to
465 evaluate HRV in cardiovascular literature. The strengths of this study consist of having accurate
466 prenatal and recent MeHg exposure measures at 19 years of age, a large cohort whose MeHg
467 exposure is significantly higher than other reported cohorts, a comprehensive battery of HRV
468 parameters completed under controlled conditions over a prolonged time period, blinding of
469 clinicians to exposure, and *a priori* specified analysis plans. Additionally, the analyses identified
470 known associations of HRV parameters indicating that if MeHg did influence autonomic control of
471 HRV the study could have detected it. As with all epidemiological studies, there are also
472 limitations. Complete data were not available on all subjects, conditions during the overnight
473 recordings were not monitored, and we were not able to complete long recordings on all
474 participants. Furthermore, our measure of recent exposure was limited to the month prior to
475 evaluation.

476 In conclusion, in a large cohort of 19 year old participants with MeHg exposures
477 significantly above those in previous reports, we found variations in HRV parameters related to
478 sex that have been previously reported, but found no association of them with prenatal MeHg
479 exposure. We found some trends toward associations of recent (at the age of 19 years) MeHg
480 exposure with heart rate and HRV parameters when subjects were challenged with math test that

481 could suggest that concurrent MeHg exposure might influence autonomic response, but the
482 magnitude of these findings is so miniscule that it could not be considered meaningful from a
483 clinical standpoint. In addition, we are cautious with this interpretation because of the multiple
484 comparisons and small magnitude of the associations. We do not believe that these data support
485 the hypothesis that either prenatal or recent MeHg exposure from consuming naturally
486 contaminated ocean fish adversely influences heart rate variability in young adults.

488 **References**

- 489 (CONTAM), EFSA Panel on Contaminants in the Food Chain. 2012. 'Scientific Opinion on the risk for public
490 health related to the presence of mercury and methylmercury in food. EFSA ', *EFSA Journal*, 10:
491 241.
- 492 Agelink, M. W., R. Malessa, B. Baumann, T. Majewski, F. Akila, T. Zeit, and D. Ziegler. 2001. 'Standardized
493 tests of heart rate variability: normal ranges obtained from 309 healthy humans, and effects of
494 age, gender, and heart rate', *Clin Auton Res*, 11: 99-108.
- 495 Antelmi, I., R. S. de Paula, A. R. Shinzato, C. A. Peres, A. J. Mansur, and C. J. Grupi. 2004. 'Influence of age,
496 gender, body mass index, and functional capacity on heart rate variability in a cohort of subjects
497 without heart disease', *Am J Cardiol*, 93: 381-5.
- 498 Bigger, J. T., Jr., J. L. Fleiss, R. C. Steinman, L. M. Rolnitzky, R. E. Kleiger, and J. N. Rottman. 1992. 'Frequency
499 domain measures of heart period variability and mortality after myocardial infarction',
500 *Circulation*, 85: 164-71.
- 501 Breitner, S., A. Peters, W. Zareba, R. Hampel, D. Oakes, J. Wiltshire, M. W. Frampton, P. K. Hopke, J. Cyrus,
502 M. J. Utell, C. Kane, A. Schneider, and D. Q. Rich. 2019. 'Ambient and controlled exposures to
503 particulate air pollution and acute changes in heart rate variability and repolarization', *Sci Rep*, 9:
504 1946.
- 505 Cernichiari, E., T. Y. Toribara, L. Liang, D. O. Marsh, M. W. Berlin, G. J. Myers, C. Cox, C. F. Shamlaye, O.
506 Choisy, P. Davidson, and et al. 1995. 'The biological monitoring of mercury in the Seychelles study',
507 *Neurotoxicology*, 16: 613-28.
- 508 Choi, A. L., P. Weihe, E. Budtz-Jorgensen, P. J. Jorgensen, J. T. Salonen, T. P. Tuomainen, K. Murata, H. P.
509 Nielsen, M. S. Petersen, J. Askham, and P. Grandjean. 2009. 'Methylmercury exposure and adverse
510 cardiovascular effects in Faroese whaling men', *Environ Health Perspect*, 117: 367-72.
- 511 Christensen, J. H., M. S. Christensen, J. Dyerberg, and E. B. Schmidt. 1999. 'Heart rate variability and fatty
512 acid content of blood cell membranes: a dose-response study with n-3 fatty acids', *Am J Clin Nutr*,
513 70: 331-7.
- 514 Clarkson, T. W., L. Magos, and G. J. Myers. 2003. 'The toxicology of mercury--current exposures and clinical
515 manifestations', *N Engl J Med*, 349: 1731-7.
- 516 Coumel, P., J. S. Hermida, B. Wennerblom, A. Leenhardt, P. Maison-Blanche, and B. Cauchemez. 1991.
517 'Heart rate variability in left ventricular hypertrophy and heart failure, and the effects of beta-
518 blockade. A non-spectral analysis of heart rate variability in the frequency domain and in the time
519 domain', *Eur Heart J*, 12: 412-22.
- 520 Cygankiewicz, I., and W. Zareba. 2013. 'Heart rate variability', *Handb Clin Neurol*, 117: 379-93.
- 521 Davidson, P. W., G. J. Myers, C. Cox, C. Axtell, C. Shamlaye, J. Sloane-Reeves, E. Cernichiari, L. Needham,
522 A. Choi, Y. Wang, M. Berlin, and T. W. Clarkson. 1998. 'Effects of prenatal and postnatal
523 methylmercury exposure from fish consumption on neurodevelopment: outcomes at 66 months
524 of age in the Seychelles Child Development Study', *JAMA*, 280: 701-7.
- 525 El Agaty, S. M., A. Kirmani, and E. Labban. 2017. 'Heart rate variability analysis during immediate recovery
526 from exercise in overweight/obese healthy young adult females', *Ann Noninvasive Electrocardiol*,
527 22.
- 528 FAO. 2016. "The State of World Fisheries and Aquaculture 2016. Contributing to food security and
529 nutrition for all. ." In, 1-200. Rome.
- 530 Fisher, J. P., C. N. Young, and P. J. Fadel. 2015. 'Autonomic adjustments to exercise in humans', *Compr*
531 *Physiol*, 5: 475-512.

532 Folch, J., M. Lees, and G. H. Sloane Stanley. 1957. 'A simple method for the isolation and purification of
533 total lipides from animal tissues', *J Biol Chem*, 226: 497-509.

534 Grandjean, P., K. Murata, E. Budtz-Jorgensen, and P. Weihe. 2004. 'Cardiac autonomic activity in
535 methylmercury neurotoxicity: 14-year follow-up of a Faroese birth cohort', *J Pediatr*, 144: 169-76.

536 Gribble, M. O., A. Cheng, R. D. Berger, L. Rosman, and E. Guallar. 2015. 'Mercury Exposure and Heart Rate
537 Variability: a Systematic Review', *Curr Environ Health Rep*, 2: 304-14.

538 Gump, B. B., M. J. Dykas, J. A. MacKenzie, A. K. Dumas, B. Hruska, C. K. Ewart, P. J. Parsons, C. D. Palmer,
539 and K. Bendinskas. 2017. 'Background lead and mercury exposures: Psychological and behavioral
540 problems in children', *Environ Res*, 158: 576-82.

541 Gump, B. B., J. A. Mackenzie, K. Bendinskas, R. Morgan, A. K. Dumas, C. D. Palmer, and P. J. Parsons. 2011.
542 'Low-level Pb and cardiovascular responses to acute stress in children: the role of cardiac
543 autonomic regulation', *Neurotoxicol Teratol*, 33: 212-9.

544 Hoogerwaard, A. F., M. R. de Jong, A. Adiyaman, J. J. J. Smit, Pphm Delnoy, J. E. Heeg, Baam van Hasselt,
545 A. R. Ramdat Misier, M. Rienstra, I. C. van Gelder, and A. Elvan. 2019. 'Renal sympathetic
546 denervation induces changes in heart rate variability and is associated with a lower sympathetic
547 tone', *Clin Res Cardiol*, 108: 22-30.

548 HRV. 1996. 'Heart rate variability. Standards of measurement, physiological interpretation, and clinical
549 use. Task Force of the European Society of Cardiology and the North American Society of Pacing
550 and Electrophysiology', *Eur Heart J*, 17: 354-81.

551 Jerling, M., I. Cygankiewicz, N. Al-Tawil, B. Darpo, A. Ljungstrom, and W. Zareba. 2018. 'Effects of intranasal
552 kinetic oscillation stimulation on heart rate variability', *Ann Noninvasive Electrocardiol*, 23.

553 Karita, K., T. Iwata, E. Maeda, M. Sakamoto, and K. Murata. 2018. 'Assessment of Cardiac Autonomic
554 Function in Relation to Methylmercury Neurotoxicity', *Toxics*, 6.

555 Kleiger, R. E., J. P. Miller, J. T. Bigger, Jr., and A. J. Moss. 1987. 'Decreased heart rate variability and its
556 association with increased mortality after acute myocardial infarction', *Am J Cardiol*, 59: 256-62.

557 Kleiger, R. E., P. K. Stein, and J. T. Bigger, Jr. 2005. 'Heart rate variability: measurement and clinical utility',
558 *Ann Noninvasive Electrocardiol*, 10: 88-101.

559 Lim, S., H. U. Chung, and D. Paek. 2010. 'Low dose mercury and heart rate variability among community
560 residents nearby to an industrial complex in Korea', *Neurotoxicology*, 31: 10-6.

561 Magos, L., and T. W. Clarkson. 1972. 'Atomic absorption determination of total, inorganic, and organic
562 mercury in blood', *J Assoc Off Anal Chem*, 55: 966-71.

563 Malik, M., T. Farrell, T. Cripps, and A. J. Camm. 1989. 'Heart rate variability in relation to prognosis after
564 myocardial infarction: selection of optimal processing techniques', *Eur Heart J*, 10: 1060-74.

565 Marsh, D. O., T. W. Clarkson, G. J. Myers, P. W. Davidson, C. Cox, E. Cernichiari, M. A. Tanner, W. Lednar,
566 C. Shamlaye, O. Choisy, and et al. 1995. 'The Seychelles study of fetal methylmercury exposure
567 and child development: introduction', *Neurotoxicology*, 16: 583-96.

568 Melo, H. M., T. C. Martins, L. M. Nascimento, A. A. Hoeller, R. Walz, and E. Takase. 2018. 'Ultra-short heart
569 rate variability recording reliability: The effect of controlled paced breathing', *Ann Noninvasive
570 Electrocardiol*, 23: e12565.

571 Miller, C., R. Karimi, S. Silbernagel, D. Kostrubiak, F. Schiavone, Q. Zhang, J. Yang, E. Rashba, and J. R.
572 Meliker. 2018. 'Mercury, omega-3 fatty acids, and seafood intake are not associated with heart
573 rate variability or QT interval', *Arch Environ Occup Health*, 73: 251-57.

574 Mozaffarian, D. 2009. 'Fish, mercury, selenium and cardiovascular risk: current evidence and unanswered
575 questions', *Int J Environ Res Public Health*, 6: 1894-916.

576 Murata, K., P. Weihe, E. Budtz-Jorgensen, P. J. Jorgensen, and P. Grandjean. 2004. 'Delayed brainstem
577 auditory evoked potential latencies in 14-year-old children exposed to methylmercury', *J Pediatr*,
578 144: 177-83.

579 Myers, G. J., P. W. Davidson, C. Cox, C. F. Shamlaye, D. Palumbo, E. Cernichiari, J. Sloane-Reeves, G. E.
580 Wilding, J. Kost, L. S. Huang, and T. W. Clarkson. 2003. 'Prenatal methylmercury exposure from
581 ocean fish consumption in the Seychelles child development study', *Lancet*, 361: 1686-92.

582 Myers, G. J., S. W. Thurston, A. T. Pearson, P. W. Davidson, C. Cox, C. F. Shamlaye, E. Cernichiari, and T. W.
583 Clarkson. 2009. 'Postnatal exposure to methyl mercury from fish consumption: a review and new
584 data from the Seychelles Child Development Study', *Neurotoxicology*, 30: 338-49.

585 Oka, T., M. Matsukura, M. Okamoto, N. Harada, T. Kitano, T. Miike, and M. Futatsuka. 2002. 'Autonomic
586 nervous functions in fetal type Minamata disease patients: assessment of heart rate variability',
587 *Tohoku J Exp Med*, 198: 215-21.

588 Periard, D., B. Beqiraj, D. Hayoz, B. Viswanathan, K. Evans, S. W. Thurston, P. W. Davidson, G. J. Myers,
589 and P. Bovet. 2015. 'Associations of baroreflex sensitivity, heart rate variability, and initial
590 orthostatic hypotension with prenatal and recent postnatal methylmercury exposure in the
591 Seychelles Child Development Study at age 19 years', *Int J Environ Res Public Health*, 12: 3395-
592 405.

593 Petersen, K. K., H. H. Andersen, M. Tsukamoto, L. Tracy, J. Koenig, and L. Arendt-Nielsen. 2018. 'The effects
594 of propranolol on heart rate variability and quantitative, mechanistic, pain profiling: a randomized
595 placebo-controlled crossover study', *Scand J Pain*, 18: 479-89.

596 Roman, H. A., T. L. Walsh, B. A. Coull, E. Dewailly, E. Guallar, D. Hattis, K. Marien, J. Schwartz, A. H. Stern,
597 J. K. Virtanen, and G. Rice. 2011. 'Evaluation of the cardiovascular effects of methylmercury
598 exposures: current evidence supports development of a dose-response function for regulatory
599 benefits analysis', *Environ Health Perspect*, 119: 607-14.

600 Shaffer, F., and J. P. Ginsberg. 2017. 'An Overview of Heart Rate Variability Metrics and Norms', *Front*
601 *Public Health*, 5: 258.

602 Shamlaye, C. F., D. O. Marsh, G. J. Myers, C. Cox, P. W. Davidson, O. Choisy, E. Cernichiari, A. Choi, M. A.
603 Tanner, and T. W. Clarkson. 1995. 'The Seychelles child development study on
604 neurodevelopmental outcomes in children following in utero exposure to methylmercury from a
605 maternal fish diet: background and demographics', *Neurotoxicology*, 16: 597-612.

606 Sherazi, S., V. Kutyifa, S. McNitt, M. K. Aktas, J. P. Couderc, B. Peterson, P. E. Bloch Thomsen, J. Kautzner,
607 A. J. Moss, and W. Zareba. 2015. 'Prognostic Significance of Heart Rate Variability Among Patients
608 Treated With Cardiac Resynchronization Therapy: MADIT-CRT (Multicenter Automatic
609 Defibrillator Implantation Trial-Cardiac Resynchronization Therapy)', *JACC Clin Electrophysiol*, 1:
610 74-80.

611 Simula, S., T. P. Laitinen, T. M. Laitinen, P. Hartikainen, and J. E. K. Hartikainen. 2017. 'Sequence of
612 cardiovascular autonomic alterations after fingolimod initiation', *Ann Noninvasive Electrocardiol*,
613 22.

614 Sorensen, N., K. Murata, E. Budtz-Jorgensen, P. Weihe, and P. Grandjean. 1999. 'Prenatal methylmercury
615 exposure as a cardiovascular risk factor at seven years of age', *Epidemiology*, 10: 370-5.

616 Srinivasan, K., S. Sucharita, and M. Vaz. 2002. 'Effect of standing on short term heart rate variability across
617 age', *Clin Physiol Funct Imaging*, 22: 404-8.

618 Stapelberg, N. J. C., D. L. Neumann, D. H. K. Shum, H. McConnell, and I. Hamilton-Craig. 2018. 'The
619 sensitivity of 38 heart rate variability measures to the addition of artifact in human and artificial
620 24-hr cardiac recordings', *Ann Noninvasive Electrocardiol*, 23.

621 Stein, P. K., K. E. Freedland, J. A. Skala, R. M. Carney, V. Davila-Roman, M. W. Rich, and R. E. Kleiger. 1997.
622 'Heart rate variability is independent of age, gender, and race in congestive heart failure with a
623 recent acute exacerbation', *Am J Cardiol*, 79: 511-2.

624 Tsuji, H., M. G. Larson, F. J. Venditti, Jr., E. S. Manders, J. C. Evans, C. L. Feldman, and D. Levy. 1996. 'Impact
625 of reduced heart rate variability on risk for cardiac events. The Framingham Heart Study',
626 *Circulation*, 94: 2850-5.

627 Umetani, K., D. H. Singer, R. McCraty, and M. Atkinson. 1998. 'Twenty-four hour time domain heart rate
628 variability and heart rate: relations to age and gender over nine decades', *J Am Coll Cardiol*, 31:
629 593-601.

630 Valera, B., E. Dewailly, and P. Poirier. 2008. 'Cardiac autonomic activity and blood pressure among Nunavik
631 Inuit adults exposed to environmental mercury: a cross-sectional study', *Environ Health*, 7: 29.

632 Valera, B., E. Dewailly, P. Poirier, E. Counil, and E. Suhas. 2011. 'Influence of mercury exposure on blood
633 pressure, resting heart rate and heart rate variability in French Polynesians: a cross-sectional
634 study', *Environ Health*, 10: 99.

635 van Wijngaarden, E., S. W. Thurston, G. J. Myers, J. J. Strain, B. Weiss, T. Zarcone, G. E. Watson, G. Zareba,
636 E. M. McSorley, M. S. Mulhern, A. J. Yeates, J. Henderson, J. Gedeon, C. F. Shamlaye, and P. W.
637 Davidson. 2013. 'Prenatal methyl mercury exposure in relation to neurodevelopment and
638 behavior at 19 years of age in the Seychelles Child Development Study', *Neurotoxicol Teratol*, 39:
639 19-25.

640 Weisberg, S. 2005. *Applied Linear Regression*. (John Wiley & Sons, Inc.: Hoboken, NJ).

641 WHO. 2007. "Exposure to mercury: A major public health concern. ." In *Preventing disease through healthy*
642 *environments*. Geneva, Switzerland: Public Health and Environment, World Health Organization.

643 Yaginuma-Sakurai, K., K. Murata, M. Shimada, K. Nakai, N. Kurokawa, S. Kameo, and H. Satoh. 2010.
644 'Intervention study on cardiac autonomic nervous effects of methylmercury from seafood',
645 *Neurotoxicol Teratol*, 32: 240-5.

646

Table 1: Characteristics of 19-year old subjects with comparison of males and females.

	All N	Mean ± SD	Females N	Mean ± SD	Males N	Mean ± SD	P value
BMI (kg/m ²)	507	22.25 ± 4.83	270	22.93 ± 5.60	237	21.47 ± 3.6	0.06
Child's Birth Weight (Kg)	514	3.18 ± 0.5	273	3.12 ± 0.48	241	3.25 ± 0.51	0.01
Prenatal MeHg (ppm)	514	6.92 ± 4.54	273	7.06 ± 4.68	241	6.77 ± 4.37	0.65
Recent MeHg (ppm)	451	10.21 ± 5.79	253	8.67 ± 4.76	198	12.16 ± 6.37	<0.01
n-3 LCPUFA (mg/ml)	482	0.05 ± 0.02	256	0.05 ± 0.02	226	0.04 ± 0.02	0.48
n-6 LCPUFA (mg/ml)	482	0.15 ± 0.04	256	0.15 ± 0.04	226	0.15 ± 0.05	0.85
Activity Level (total MET):	N (%)		N (%)		N (%)		
Sedentary (< 600)	171 (33.6%)		137 (50.6%)		34 (14.3%)		<0.01
Moderate (600 - < 3,000)	139 (27.3%)		87 (32.1%)		52 (21.9%)		0.01
High (3,000+)	199 (39.1%)		47 (17.3%)		152 (63.9%)		<0.01

Activity level: categorized from participants reports of their weekly walking, working, and recreational activities, in minutes estimated time per week (MET).

P values are from the nonparametric Wilcoxon rank-sum test

n-3 = sum of DHA, EPA, and ALA. n-6 = sum of AA and LA

Table 2. Average heart rate variability measurements by categories of prenatal and recent (at 19 years of age) MeHg (ppm) exposure[^]

Outcome	Prenatal MeHg			P value	Recent MeHg			P value*
	0-<5 ppm (n=211) Mean±SD	5-<10 ppm (n=184) Mean±SD	10+ ppm (n=119) Mean±SD		0-<5 ppm (n=63) mean±SD	5-<10 ppm (n=201) mean±SD	10+ ppm (n=187) mean±SD	
NN (ms)								
Long-term Recording	794±84	795±95	825±98	0.19	768±75	806±90	806±102	0.12
Baseline Supine	928±167	915±162	922±154	0.76	890±156	907±160	936±168	0.08
Response to Standing Δ	157±99	149±94	160±92	0.43	152±99	141±93	168±96	0.02
Response to Math Test Δ	65±72	67±80	81±87	0.28	78±84	59±71	80±87	0.04
Response to Sleep Δ	-101±153	-110±146	-140±114	0.44	-112±130	-109±150	-118±142	0.98
SDNN (ms)								
Long-term Recording	181±40	181±421	182±39	0.97	173±44	180±41	184±40	0.31
Baseline Supine	83±39	80±37	82±36	0.74	83±39	78±34	86±41	0.39
Response to Standing Δ	5±33	8±32	3±27	0.73	12±32	3±27	8±37	0.36
Response to Math Test Δ	9±24	8±27	6±28	0.74	10±28	5±23	11±28	0.20
Response to Sleep Δ	-11±40	-17±43	-17±38	0.64	-4±34	-12±38	-18±46	0.29
rMSSD (ms)								
Long-term Recording	66±28	63±33	64±24	0.52	64±25	63±25	69±35	0.88
Baseline Supine	77±42	73±44	74±42	0.46	76±44	71±41	79±46	0.37
Response to Standing Δ	31±32	31±35	31±32	0.70	35±38	27±29	35±35	0.13
Response to Math Test Δ	11±27	12±30	11±34	0.49	14±35	8±24	15±36	0.25
Response to Sleep Δ	-10±36	-11±40	-18±32	0.60	-8±36	-15±36	-9±41	0.44
LF								
Baseline Supine	39±16	41±17	40±16	0.72	35±15	42±17	39±15	0.03
Response to Standing Δ	-15±25	-15±23	-13±26	0.72	-17±24	-12±25	-17±24	0.23
Response to Math Test Δ	-7±19	-4±22	-6±20	0.51	-8±19	-3±22	-8±18	0.23
Response to Sleep Δ	3±22	8±21	8±22	0.33	0±18	10±22	3±23	0.03
HF								
Baseline Supine	50±19	49±20	40±16	0.72	55±18	47±19	51±19	0.01
Response to Standing Δ	29±21	26±21	-13±26	0.72	30±24	23±21	30±20	0.01
Response to Math Test Δ	7±19	7±22	-6±20	0.51	10±20	4±20	10±20	0.10
Response to Sleep Δ	-4±26	-3±27	8±22	0.33	0±27	-10±27	1±24	0.02

[^]P = value is from 2-df nonparametric Kruskal Wallis ANOVA test

Δ = Difference between baseline values and values obtained at standing, math test, or sleeping, respectively

* Please note that significant differences were observed when comparing middle MeHg dose group with lower and higher dose groups without dose-dependent effect.

Table 3. Prenatal MeHg (ppm) exposure association with heart rate variability measurements, adjusting for sex, birth weight and BMI

Outcome	N	P value for sex interaction	β	P value	95% CI
NN (ms)					
Long-term Recording	202	0.0230	1.883	0.1160	(-0.4710, 4.2360)
female			-0.1729	0.9073	(-3.0974, 2.7516)
male			5.494	0.0058	(1.6105, 9.3775)
Baseline Supine	507	0.3050	-1.303	0.3470	(-4.0210, 1.4150)
Response to Standing Δ	497	0.2717	-0.185	0.8420	(-2.0120, 1.6410)
Response to Math Test Δ	442	0.5480	1.534	0.0620	(-0.0760, 3.1440)
Response to Sleep Δ	202	0.0120	-2.915	0.2000	(-7.3900, 1.5600)
female			1.3949	0.6203	(-4.1492, 6.9390)
male			-10.4873	0.0055	(-17.8493, -3.1253)
SDNN (ms)					
Long-term Recording +	200	0.1820	-0.001	0.7160	(-0.0080, 0.0050)
Baseline Supine +	507	0.4077	-0.004	0.4120	(-0.0120, 0.0050)
Response to Standing *	497	0.8190	-0.001	0.7080	(-0.0090, 0.0060)
Response to Math Test *	442	0.9039	0.000	0.9360	(-0.0070, 0.0060)
Response to Sleep *	202	0.7592	-0.003	0.7310	(-0.0180, 0.0130)
rMSSD (ms)					
Long-term Recording +	202	0.5522	-0.006	0.3470	(-0.0180, 0.0060)
Baseline Supine +	499	0.2371	-0.008	0.1730	(-0.0190, 0.0030)
Response to Standing *	490	0.3928	0.002	0.6740	(-0.0070, 0.0110)
Response to Math Test *	434	0.3294	0.003	0.3760	(-0.0040, 0.0110)
Response to Sleep *	199	0.6891	-0.001	0.9150	(-0.0170, 0.0150)
LF					
Baseline Supine	505	0.9939	0.07	0.6610	(-0.2440, 0.3850)
Response to Standing Δ	496	0.3730	0.037	0.8800	(-0.4390, 0.5120)
Response to Math Test Δ	441	0.7007	-0.05	0.8130	(-0.4620, 0.3630)
Response to Sleep Δ	201	0.6549	0.246	0.4730	(-0.4280, 0.9200)
HF					
Baseline Supine	505	0.4400	-0.064	0.7310	(-0.4300, 0.3020)
Response to Standing Δ	496	0.3806	-0.12	0.5650	(-0.5310, 0.2910)
Response to Math Test Δ	441	0.2875	0.131	0.5230	(-0.2700, 0.5320)
Response to Sleep Δ	201	0.1498	-0.107	0.8010	(-0.9380, 0.7250)

+ = Model uses the natural logarithm of the outcome, and slopes are multiplicative effects

Δ = Difference between baseline values and values obtained at standing, math test, or sleeping respectively

* = Difference in natural logarithms of baseline values and values obtained at standing, math test, or sleeping, respectively

If Male*MeHg interaction is NOT significant ($p > .05$), mercury coefficient reported for entire sample

If Male*MeHg interaction IS significant ($p \leq .05$), mercury coefficient reported by sex

Table 4. Recent MeHg (ppm) exposure and n-3 and n-6 PUFA status association with heart rate variability measurements, adjusting for birth weight, BMI, and activity level

Outcome	N	P value for sex interaction	β	P value	95% CI
NN (ms)					
Long-term Recording	167	0.3404	-2.755	0.0270	(-5.191, -0.319)
Baseline Supine	418	0.0717	-3.048	0.0160	(-5.525, -0.571)
Response to Standing Δ	411	0.0185	0.167	0.8420	(-1.485, 1.82)
female			2.617	0.0499	(0.0011, 5.2323)
male			-1.426	0.1847	(-3.5366, 0.6838)
Response to Math Test Δ	365	0.1291	0.578	0.4420	(-0.9, 2.056)
Response to Sleep Δ	167	0.4733	-0.095	0.9680	(-4.77, 4.581)
SDNN (ms)					
Long-term Recording +	165	0.4254	-0.006	0.0950	(-0.012, 0.001)
Baseline Supine +	418	0.6280	-0.005	0.1870	(-0.013, 0.003)
Response to Standing Δ^*	411	0.5215	0	0.9400	(-0.007, 0.007)
Response to Math Test Δ^*	365	0.7105	0.004	0.1480	(-0.002, 0.01)
Response to Sleep Δ^*	167	0.4633	-0.010	0.2050	(-0.026, 0.006)
rMSSD (ms)					
Long-term Recording +	167	0.1793	-0.010	0.1300	(-0.022, 0.003)
Baseline Supine +	411	0.5038	-0.008	0.1280	(-0.018, 0.002)
Response to Standing Δ^*	405	0.1855	0.004	0.3360	(-0.004, 0.012)
Response to Math Test Δ^*	358	0.4383	0.010	0.0060	(0.003, 0.016)
Response to Sleep Δ^*	165	0.4679	-0.007	0.4200	(-0.024, 0.01)
LF					
Baseline Supine	416	0.3200	-0.128	0.3860	(-0.417, 0.161)
Response to Standing Δ	410	0.2926	-0.361	0.1060	(-0.798, 0.077)
Response to Math Test Δ	364	0.0425	-0.245	0.1910	(-0.612, 0.122)
female			0.209	0.4720	(-0.3615, 0.7791)
male			-0.563	0.0210	(-1.0401, -0.0855)
Response to Sleep Δ	166	0.2318	-0.688	0.0500	(-1.377, 0.001)
HF					
Baseline Supine	416	0.6151	0.226	0.1890	(-0.112, 0.564)
Response to Standing Δ	410	0.8996	0.490	0.0110	(0.114, 0.866)
Response to Math Test Δ	364	0.7939	0.544	0.0030	(0.188, 0.901)
Response to Sleep Δ	166	0.8533	0.814	0.0550	(-0.018, 1.646)

+ = Model uses the natural logarithm of the outcome and slopes assume multiplicative effects

Δ = Difference between baseline values and values obtained at standing, math test, or sleeping respectively

* = Difference in natural logarithms of baseline values and values obtained at standing, math test, or sleeping respectively

If Male*MeHg interaction is not significant ($p > .05$), mercury coefficient reported for entire sample

If Male*MeHg interaction is significant ($p \leq .05$), the mercury coefficient is reported by sex

Supplementary Tables

For the manuscript entitled: "Prenatal and Recent Methylmercury Exposure and Heart Rate Variability in Young Adults: the Seychelles Child Development Study" by Zareba et al.

Table S1: Heart rate variability parameters (n=23) from Holter recordings comparing males and females.

	All		Female		Male		P value
	N	Mean ± SD	N	Mean ± SD	N	Mean ± SD	
NN (ms)							
Long-term Recording	203	802 ± 92	117	757 ± 68	86	863 ± 85	<0.0001
Baseline Supine	514	922 ± 162	273	847 ± 122	241	1007 ± 161	<0.0001
Response to Standing Δ	504	155 ± 95	267	133 ± 85	237	179 ± 101	<0.0001
Response to Math Test Δ	448	69 ± 79	240	69 ± 79	208	70 ± 80	0.9731
Response to Sleep Δ	203	-113 ± 142	117	-102 ± 114	86	-128 ± 173	0.0414
SDNN (ms)							
Long-term Recording	201	181 ± 40	117	166 ± 35	84	203 ± 36	<0.0001
Baseline Supine	514	82 ± 38	273	74 ± 37	241	90 ± 36	<0.0001
Response to Standing Δ	504	6 ± 32	267	8 ± 31	237	3 ± 33	0.0951
Response to Math Test Δ	448	8 ± 26	240	9 ± 24	208	6 ± 28	0.2266
Response to Sleep Δ	203	-14 ± 40	117	-11 ± 32	86	-19 ± 50	0.3927
rMSSD (ms)							
Long-term Recording	203	65 ± 28	117	59 ± 25	86	72 ± 30	0.0002
Baseline Supine	506	75 ± 43	267	67 ± 42	239	83 ± 42	<0.0001
Response to Standing Δ	497	31 ± 33	262	30 ± 33	235	33 ± 33	0.0217
Response to Math Test Δ	440	11 ± 30	234	14 ± 28	206	8 ± 32	0.0149
Response to Sleep Δ	200	-12 ± 36	116	-13 ± 34	84	-11 ± 40	0.9733
LF							
Baseline Supine	512	40 ± 16	272	38 ± 16	240	42 ± 16	0.0119
Response to Standing Δ	503	-15 ± 24	266	-13 ± 25	237	-17 ± 24	0.1223
Response to Math Test Δ	447	-6 ± 20	239	-5 ± 21	208	-6 ± 20	0.8824
Response to Sleep Δ	202	6 ± 22	117	3 ± 22	85	10 ± 21	0.0131
HF							
Baseline Supine	512	49 ± 19	272	51 ± 20	240	48 ± 18	0.0774
Response to Standing Δ	503	27 ± 21	266	29 ± 22	237	25 ± 20	0.0415
Response to Math Test Δ	447	7 ± 20	239	8 ± 21	208	6 ± 19	0.0911
Response to Sleep Δ	202	-4.7 ± 27	117	-3 ± 28	85	-7 ± 25	0.2488

Δ = Difference between baseline values and values obtained at standing, math test, or sleeping, respectively.

P values are from the nonparametric Wilcoxon rank-sum test

Table S2. Time-domain HRV parameters from long-term ECG recordings according to recent MeHg level categories at 19 years of age in males and females separately.

Recent MeHg Levels:	0-5 ppm	5-10 ppm	10+ppm	P value
Males	N=14	N=77	n=109	
NN	899 ± 147	866 ± 121	871 ± 122	0.66
SDNN	187 ± 50	181 ± 49	183 ± 45	0.88
rMSSD	84 ± 37	77 ± 29	74 ± 37	0.31
pNN50	16.9 ± 6.7	13.8 ± 6.9	13.8 ± 6.1	0.24
SDANN	148 ± 43	153 ± 54	155 ± 48	0.90
SDNNIX	108 ± 36	93 ± 26	95 ± 27	0.19
Females	n=49	n=127	n=79	
NN	752 ± 91	759 ± 82	758 ± 82	0.87
SDNN	150 ± 44	144 ± 45	143 ± 41	0.71
rMSSD	60 ± 27	57 ± 29	64 ± 38	0.32
pNN50	11.2 ± 6.2	11.0 ± 6.7	11.2 ± 6.9	0.96
SDANN	126 ± 46	119 ± 42	116 ± 40	0.45
SDNNIX	75 ± 22	74 ± 23	77 ± 27	0.65

The same findings of no significant difference among three groups were observed for all other tested HRV parameters (data not shown - could be provided if needed).

Table S3. Recent MeHg (ppm) exposure association with heart rate variability measurements after adjusting for sex, birth weight and BMI (Primary Models)

Outcome	N	P value for sex interaction	β	P value	95% CI
NN (ms)					
Long-term Recording	174	0.3674	-1.821	0.1110	(-4.0650, 0.4230)
Baseline Supine	446	0.0975	-1.448	0.2370	(-3.8540, 0.9570)
Response to Standing Δ	438	0.0149	0.645	0.4250	(-0.9420, 2.2330)
AC.RR:female			2.9957	0.0172	(0.5341, 5.4573)
AC.RR:male			-0.9796	0.3479	(-3.0282, 1.069)
Response to Math Test Δ	391	0.0547	0.646	0.3720	(-0.7760, 2.0680)
Response to Sleep Δ	174	0.4293	0.512	0.8120	(-3.7330, 4.7570)
SDNN (ms)					
Long-term Recording +	172	0.2179	-0.005	0.1280	(-0.0110, 0.0010)
Baseline Supine +	446	0.5166	-0.002	0.6000	(-0.0100, 0.0060)
Response to Standing *	438	0.4297	0.001	0.8650	(-0.0060, 0.0070)
Response to Math Test *	391	0.8776	0.005	0.1150	(-0.0010, 0.0100)
Response to Sleep *	174	0.7759	-0.002	0.8050	(-0.0170, 0.0130)
rMSSD (ms)					
Long-term Recording +	174	0.1906	-0.008	0.1970	(-0.0190, 0.0040)
Baseline Supine +	438	0.4000	-0.003	0.4890	(-0.0130, 0.0060)
Response to Standing *	431	0.0783	0.005	0.2250	(-0.0030, 0.0130)
Response to Math Test *	383	0.1257	0.009	0.0070	(0.0020, 0.0160)
Response to Sleep *	171	0.9234	0.003	0.7520	(-0.0130, 0.0180)
LF					
Baseline Supine	444	0.4601	-0.065	0.6390	(-0.3400, 0.2090)
Response to Standing Δ	437	0.5816	-0.323	0.1260	(-0.7360, 0.0910)
Response to Math Test Δ	390	0.0940	-0.162	0.3660	(-0.5130, 0.1900)
Response to Sleep Δ	173	0.0929	-0.58	0.0730	(-1.2140, 0.0540)
HF					
Baseline Supine	444	0.7396	0.188	0.2510	(-0.1340, 0.5100)
Response to Standing Δ	437	0.9000	0.403	0.0290	(0.0420, 0.7650)
Response to Math Test Δ	390	0.9269	0.375	0.0320	(0.0330, 0.7160)
Response to Sleep Δ	173	0.5599	0.597	0.1270	(-0.1710, 1.3650)

+ = Model uses the natural logarithm of the outcome and slopes are multiplicative effects

Δ = Difference between baseline values and values obtained at standing, math test, or sleeping, respectively

* = Difference in natural logarithms of baseline values and values obtained at standing, math test, or sleeping, respectively. If Male*MeHg interaction is NOT significant ($p > .05$), mercury coefficient reported for entire sample
If Male*MeHg interaction IS significant ($p \leq .05$), mercury coefficient reported by sex