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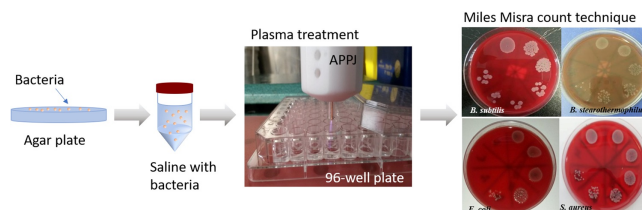
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Abstract Atmospheric pressure plasmas are becoming relevant in local microbial deactivation and other combined effects of plasmas on living organisms. For this reason, our research was focussed on optimisation of atmospheric pressure plasma jet (APPJ) parameters to complete the deactivation of different bacteria strains in a medium. Different helium APPJ treatments with different discharge parameters were used, such as input voltages and gas flows. To better understand plasma properties behind complete bacteria deactivation at optimised discharge parameters, optical and electrical plasma jet diagnostics were performed, including electrical characterisation of the plasma source, optical emission spectroscopy of the plasma plume and intensified charged coupled device imaging of the discharge behaviour for every set of plasma parameters. Then, the resulting plasma liquid chemistry was assessed to establish the connections between reactive species generated in the gaseous and liquid phases. The most efficient deactivation was found for higher discharge powers and gas flow rates, and that was linked to higher densities of reactive oxygen and nitrogen species, especially hydrogen peroxide and medium solvated charges.

Graphical abstract:





Helium atmospheric pressure plasma jet parameters and their influence on bacteria deactivation in a medium

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Abstract. Atmospheric pressure plasmas are becoming relevant in local microbial deactivation and other combined effects of plasmas on living organisms. For this reason, our research was focussed on optimisation of atmospheric pressure plasma jet (APPJ) parameters to complete the deactivation of different bacteria strains in a medium. Different helium APPJ treatments with different discharge parameters were used, such as input voltages and gas flows. To better understand plasma properties behind complete bacteria deactivation at optimised discharge parameters, optical and electrical plasma jet diagnostics were performed, including electrical characterisation of the plasma source, optical emission spectroscopy of the plasma plume and intensified charged coupled device imaging of the discharge behaviour for every set of plasma parameters. Then, the resulting plasma liquid chemistry was assessed to establish the connections between reactive species generated in the gaseous and liquid phases. The most efficient deactivation was found for higher discharge powers and gas flow rates, and that was linked to higher densities of reactive oxygen and nitrogen species, especially hydrogen peroxide and medium solvated charges.

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1 Introduction

It is well known that some microorganisms, such as bacteria, fungi and viruses, act as pathogens and induce various diseases. Moreover, microorganisms can cause food spoilage and damage to materials such as corrosion of plumbing systems. For these reasons, several conventional sterilisation techniques which lead to complete microbial deactivation or removal have been developed, including heating, filtration, chemical liquid agents and radiation. However, a disadvantage of these sterilisation techniques is that they can be used only on thermally resistant and chemically inert substrates, as those techniques can influence substrate properties [1].

In recent years, non-thermal atmospheric pressure plasmas have been proposed as an alternative to conventional sterilisation techniques. Most frequently reported is sterilisation with atmospheric pressure plasma jets (APPJs) due to their low operating temperatures and cost-effective operation [2–5]. APPJs are suitable for selective treatment of specific substrates as they contain more known inactivation agents without

the downsides of conventional sterilisation techniques. Research suggests that reactive oxygen species play the biggest role in bacteria inactivation, but UV radiation, electric field, other reactive species and charged particles also contribute to the process [5–8]. In this way, APPJs represent one of the most promising discharge candidates for different biological applications, including complete deactivation of bacteria [9–12].

This research tested the efficiency of a constructed APPJ on four different bacteria: *Escherichia coli*, *Staphylococcus aureus*, *Bacillus subtilis*, and *Bacillus stearothermophilus*. *B. stearothermophilus* and *B. subtilis* are spore-forming bacteria and the most commonly recognised and widely used biological indicators for monitoring the effectiveness of sterilisation processes. Spores are dormant bacterial structures, highly resistant to disinfectants and sterilising agents. Spore-forming bacteria are commonly found in processed foods and dairy products [13–15]. These bacteria were tested in order to see how an APPJ affects spore-forming bacteria. Additionally, *E. coli* and *S. aureus*, the most common pathogens in humans and widespread in nature (in hospitals and working and living surroundings), were selected. They are commonly found

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in different environments, contaminating various items, medical tools and food, and can cause hospital infections and food poisoning, as well as medically severe and sometimes fatal infections [16–19]. Moreover, these bacterial strains are known to possibly be multidrug-resistant [20, 21]. In addition, these bacteria tend to form biofilms where bacteria are well protected from the outside agents. It has been shown that plasmas may sterilize even the biofilms as well as planktonic samples [22].

There have been many reports on atmospheric pressure plasma-induced bacteria deactivation and decontamination [23–27]. However, there is a knowledge gap in optimising plasma parameters so that complete bacteria deactivation in a medium can be achieved in the shortest (optimal) times. Therefore, this research focuses on finding the most efficient parameters of a non-thermal helium APPJ as one of the most frequently used sources for deactivating bacteria. For this purpose, various combinations of input DC power unit voltages and gas flows were tested. Appropriate diagnostics were done both on plasma source and discharge, and on medium bacteria were suspended in.

2 Experimental setup

2.1 Preparation of bacteria samples

The deactivation effect of an APPJ, operated with helium as a working gas, was investigated on four different types of bacteria: *B. stearothermophilus* (ATCC No. 7953), *B. subtilis* (ATCC No. 6633), *S. aureus* (ATCC No. 25923) and *E. coli* (ATCC No. 25922). Bacterial cultures were grown overnight on Columbia (COS) agar plates (bioMérieux SA, Marcy l’Etoile, France) at 55 °C for *B. stearothermophilus* and 37 °C for *B. subtilis*, *S. aureus* and *E. coli*. bacteria were picked up with a loop and resuspended in sterile saline to obtain 0.5 McF (1.5×10^8 CFU/ml) initial bacterial suspension. The concentration was constant in all experiments. 100 µl of these initial 0.5 McF bacterial suspensions was evenly transferred to a 96-well plate with a flat bottom. Bacterial suspensions were exposed to the He APPJ at a constant distance for different exposure times. The samples were treated each time in triplicates.

To determine viable counts and evaluate plasma treatment effects, the Miles and Misra viable count

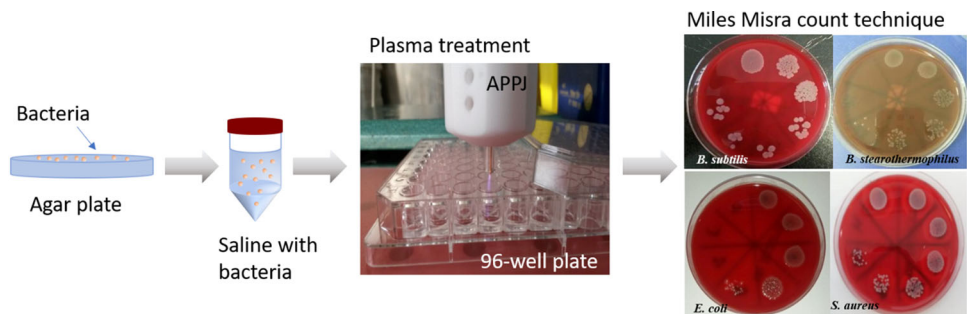
technique on COS blood agar plate (bioMérieux SA, Marcy l’Etoile, France) was used. A 20 µl properly diluted plasma-treated bacterial suspension, as well as a positive (untreated bacterial suspension) and negative control (sterile saline), was placed onto the blood agar plate. This procedure is depicted in Fig. 1. Measurements of the reactive species and pH were also conducted. Reactive species concentrations of NO_2^- and H_2O_2 were measured by a spectrophotometer (UV VIS Lambda 25) via colorimetric assays in sterile saline. The pH measurements were performed by a pH-meter (Sentron®) also in saline.

2.2 APPJ system

Bacteria-containing medium was treated by APPJ source which is designed to be handheld and highly portable, schematic of which is shown in Fig. 2. The portability of the device was facilitated by a small-size custom-made power source which is placed inside a $20 \times 12 \times 6$ cm plastic box including connectors and switches. It was connected to a commercial DC power supply Voltcraft SPS12-12 W-A. The power source output signal amplitude was varied by changing the DC input signal voltage, at discrete voltages 3, 4.5, 6, 9 and 12 V. Based on the DC input voltage, the output signal at 16 kHz was supplied to the jet enabling plasma operation at several high voltages from 1200 to 3500 V (RMS values). The jet had a copper wire, which serves as needle-type powered electrode that was placed inside the glass tube with inner diameter of 2 mm and outer diameter of 4 mm. The tube itself is held by 125 mm long and 26 mm wide Teflon housing, from which it protrudes 8 mm on the one side. On the other side, the tube was connected to gas inlet and Bronkhorst Mass-View MV-194 flow controller. For these treatments, we used He as a working gas at various fixed flows (0.5, 1, 1.5 and 2 slm).

Optical characterisation comprised optical emission spectrometry and plasma imaging by using an intensified charged coupled device (ICCD) camera. Optical emission spectroscopy was performed with an Andor Shamrock 500i spectrometer equipped with iXon Ultra 897 as a detector. An optical fibre was used to receive the emission from the plasma plume and direct it to the entrance slit of the spectrometer. The fibre was positioned at a distance of 5 cm from the jet tube

Fig. 1 Schematic representation of the experimental protocol procedure



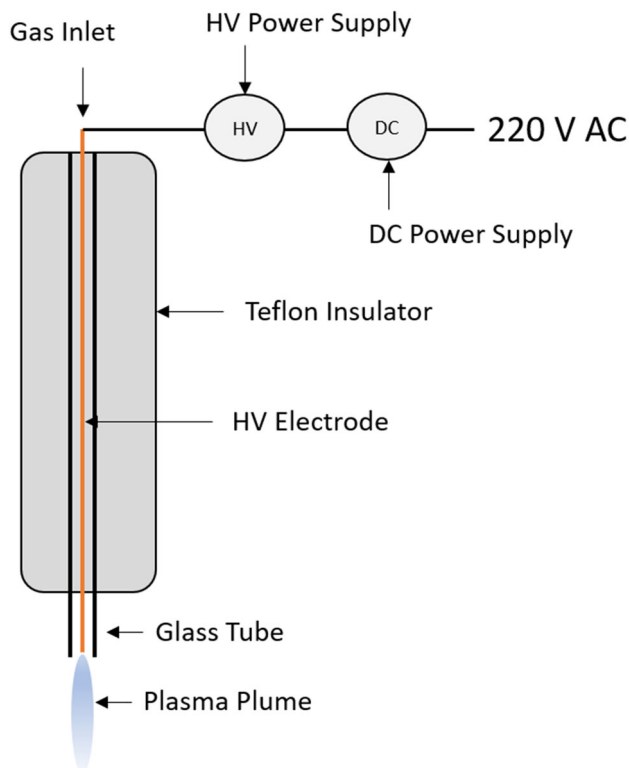


Fig. 2 Schematic representation of an APPJ used in this research

3 Results and discussion

3.1 Bacteria deactivation

At first, bacterial suspension control samples were exposed only to helium gas flow with rates of 0.5, 1, 1.5 and 2 slm, without plasma and with no voltage applied, for the same duration as required for deactivation using the plasma. The obtained results exhibit no difference in bacteria viability (Fig. 3) compared to the untreated samples (positive control; PC), which confirms that helium alone is insufficient for bacteria deactivation. The effects of the APPJ were then further tested for all bacteria and analysed with a quantitative and informative approach, which involved dynamical studies of bacterial growth after treatments. Typically, survival curves were determined as the numbers of colony-forming units (CFUs; surviving culturable bacteria as a function of plasma treatment time). However, to limit the presentation, only complete bacteria deactivation, achieving sterility of the medium, is shown in Fig. 4.

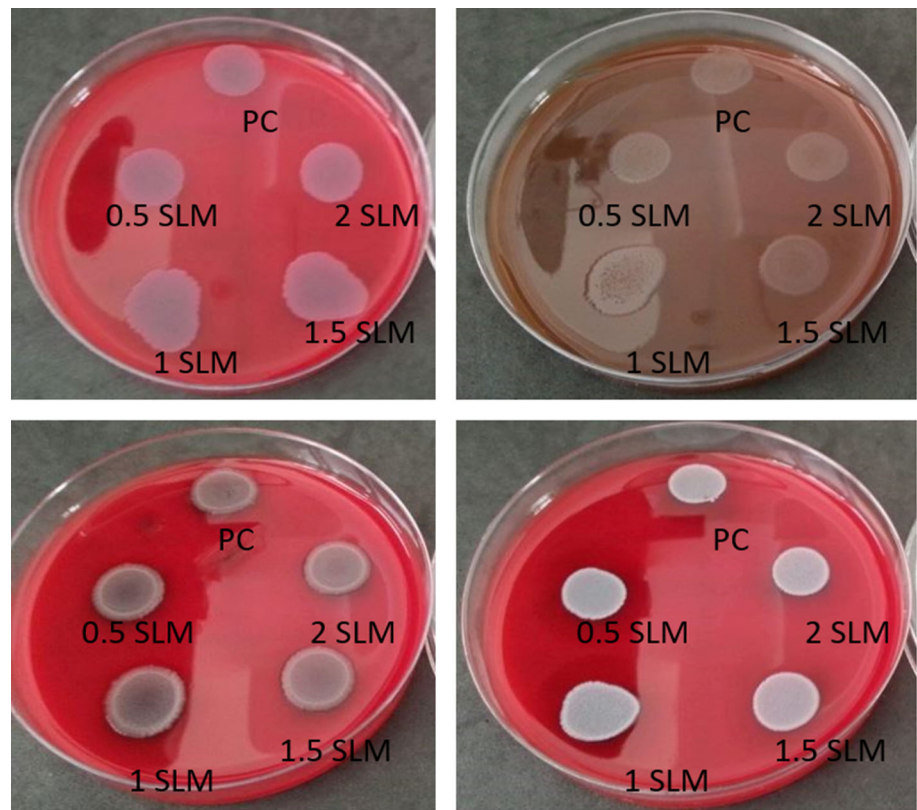
Furthermore, Fig. 4 presents the time needed for complete deactivation of *E. coli*, *S. aureus*, *B. stearothersophilus* and *B. subtilis* within a medium, exposed to a He APPJ generated with different DC input powers and gas flows. If there are no data shown for a specific set of parameters (usually 3 V and 0.5 slm), the bacteria were not completely deactivated within the maximum treatment time of 240 s used in experiments. In most cases, it was found that *E. coli* was deactivated faster than other bacteria, proving to be a less plasma-resistant strain. In this case, the highest treatment time was 180 s for deactivation under the lowest gas flow of 0.5 slm, which typically did not prove very efficient. *B. stearothersophilus* strains proved the most resistant to plasma treatments. Surprisingly, the lowest flow rate deactivation curves for 0.5 slm are very similar for all types of bacterial strains, except a small deviation with *E. coli*. This indicates that the APPJ generated at these conditions and its consequent reactive oxygen and nitrogen species (RONS) chemistry within the medium are similar (although, moving to higher flow rates, the chemistries and deactivations changed significantly). The trend follows the same patterns, where 0.5 slm is the least, and 2 slm is the most efficient, which are plasma properties connected to its subsequent interaction. The exception to this general rule is *B. stearothersophilus*, the most thermally stable and resistant strain, which seems to deviate from the rule. In this case, the most efficient chemistry for deactivation is at 1 slm. Chemical analyses of the medium chemistry elucidate the reasons for this behaviour in the following paragraphs.

From the perspective of the DC input voltage parameter used for jet discharge, the general rule is: the higher the energy input into discharge, the faster the deactivation of bacterial strains. However, it seems there is a minimum level at which the jets are efficient. It was found that an input DC voltage of 3 V was not sufficient

axis to gather the light coming from the whole channel volume. Recording of the spectra was performed for exposure times of 100 ms and with averaging of 10 spectrum acquisitions. Thus, obtained results represent space- and time-averaged emission from the plasma. Plasma imaging was performed with an Andor iStar ICCD camera DH334T-18U-03 equipped with a photographic objective. Images were taken in single-shot mode with an exposure time of 20 ms. Furthermore, electrical characterisation was performed by measuring the average power given to the jet. Voltage and current on the powered electrode were measured before the APPJ with an oscilloscope (Rigol DS1102E), high voltage probe (Rigol RP1018H) and current monitor (Pearson 8590C).

Estimation of saline solution evaporation during treatments was performed to evaluate changes in the treatment conditions throughout the experiments. For the longest treatment times, the highest DC supply voltages and He flow of 2 slm, the evaporated solution volume from the 96-well plate was not more than 50 μ l. This change in volume caused a maximum liquid level reduction of 1.2 mm, thus increasing the distance between the plasma jet and the liquid surface. However, these changes did not drastically influence plasma properties, and these maximum values were reached only for the longest treatment times and plasma powers. For most treatment conditions, volume changes fell within the experimental error of transferring the liquid volume into the plate.

Fig. 3 Effect of gas flow-only (no plasma) treatment of bacteria **a** *B. subtilis*, **b** *B. stearothermophilus*, **c** *E. coli* and **d** *S. aureus* exposed to 0.5, 1, 1.5 and 2 slm compared to the positive control (PC) by Miles and Misra plate counting



223 to deactivate most bacteria strains even for the highest
 224 gas flow and treatment times because the plasma plume
 225 was the shortest and was not in direct contact with
 226 the substrate. If deactivation of the bacteria strain was
 227 achieved, then the treatment time was significantly pro-
 228 longed. Therefore, the results indicate that He APPJ
 229 is most efficient at bacteria strain deactivation with
 230 higher applied power and higher gas flows, considering
 231 marked limits in discharge parameters and experimen-
 232 tal constraints. We do not reach conditions where addi-
 233 tional heating would produce thermal necrosis (40 °C)
 234 in the covered range of powers. While increasing effi-
 235 ciency with power is expected as for the flow, one could
 236 expect that beyond some point, further increasing of
 237 the flow may reduce efficiency by affecting the chain of
 238 plasma chemical events needed to produce the radicals
 239 that cause sterilization.

240 3.2 Chemical analysis of reactive species of saline 241 medium treated by APPJ

242 To explain the obtained results for bacterial deacti-
 243 vation in a medium, the initiated medium chemistry
 244 was investigated, determining RONS species, especially
 245 H₂O₂ and NO₂⁻ concentrations of the APPJ treated
 246 saline solution. Immediately after treatments, we per-
 247 formed measurements of pH changes. These parameters
 248 are known to influence the viability of bacterial strains
 249 significantly, as marked by numerous reports [28–31].

250 *pH measurements* were made under the same condi-
 251 tions as for reactive species measurement. The influence

of different He plasma parameters (different gas flow
 rates of 0.5, 1, 1.5 and 2 slm and different input DC
 powers of 3, 4.5, 6, 9 and 12 V) on pH value was system-
 atically measured. A pH value decrease was observed
 during the plasma treatment for most cases (Fig. 5).
 These decreasing trends featured an initial drop and
 then a steady decrease. An exception was 1 slm, which
 had an increasing pH trend for input DC voltage of 3 V.
 This could be explained by the fact that the plasma jet
 did not touch the surface of the liquid, and in this case,
 the chemistry of the medium was different than in other
 cases.

Reactive species concentrations of NO₂⁻ and H₂O₂
 were determined after plasma treatment of saline; 50 μl
 of sterile saline was placed in the 96-well plate with a
 flat bottom. The distance between the bottom of the
 well and the APPJ orifice was 15 mm, as for the treat-
 ment of bacteria, and was kept constant during the
 treatment. The results are presented in Fig. 6, and the
 results are obtained with only the parameters yield-
 ing the most efficient plasma treatment—input volt-
 age of 12 V and a flow rate of 2 slm. An expected,
 steady increase of H₂O₂ concentrations was observed
 for increasing treatment time. In contrast, the concen-
 tration of NO₂⁻ increased until 30 s, where it reached
 its maximum value and then started decreasing. The
 concentration dropped to zero after 120 s. This could
 be explained through decreasing of the pH value during
 the treatment. NO₂⁻ is very sensitive to low pH values,
 which is the cause of its decomposition or transfor-
 mation into other compounds [32].

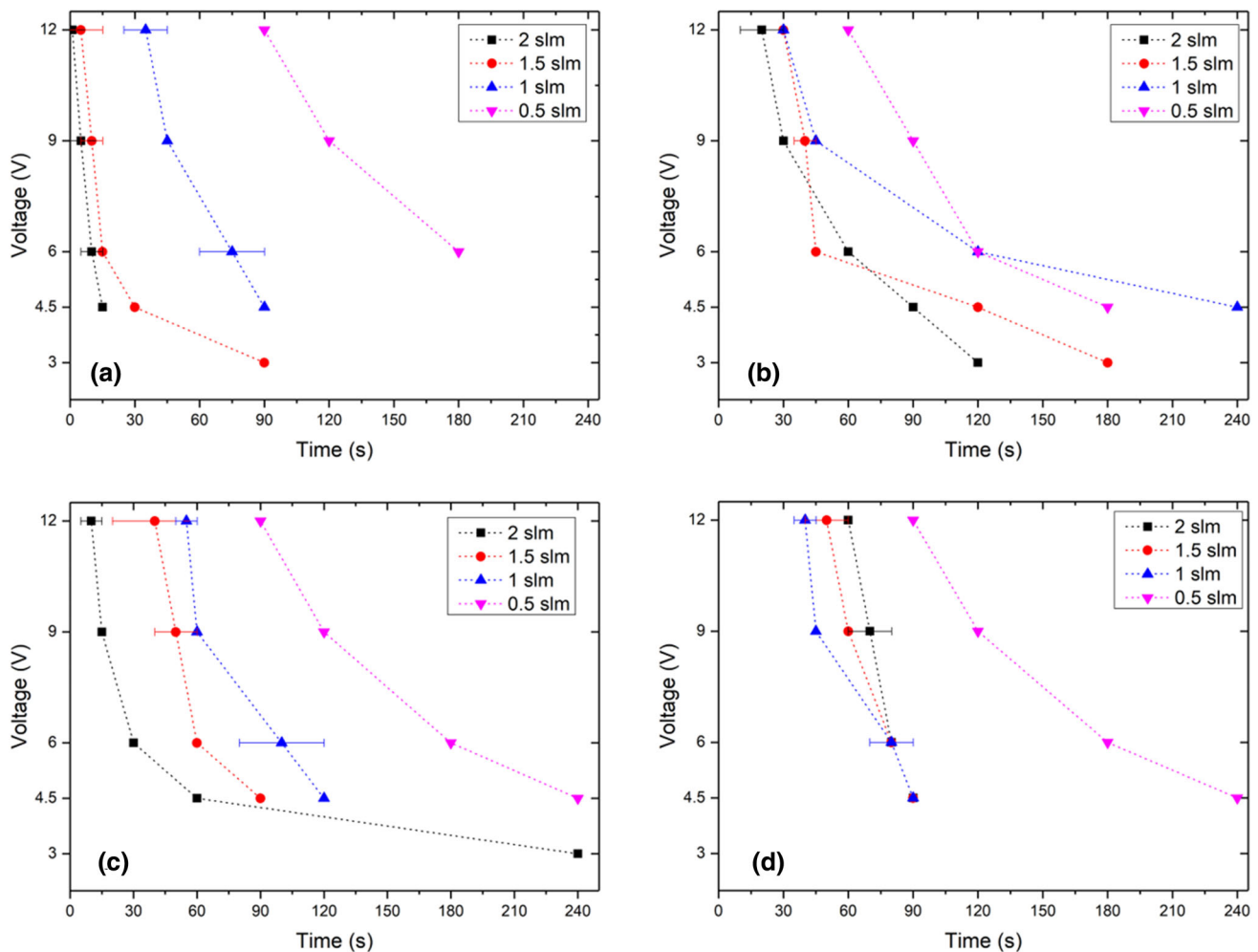


Fig. 4 Points of complete bacteria deactivation (the point at which the initial CFU/ml concentration drops to zero) **a** *E. coli*, **b** *S. aureus*, **c** *B. subtilis* and **d** *B. stearothermophilus* exposed to He APPJ generated with powers of 3, 6, 9 and 12 V, and flow rates of 0.5, 1, 1.5 and 2 slm

3.3 Diagnostics of the plasma source

In order to analyse properties of the plasma used for treatments, we performed diagnostic experiments at the same conditions as when treating media with bacterial strains. Due to safety, a saline medium was used without bacteria for these measurements.

Power measurements were made via electrical characterisation, where the average power (P_{avg}) input into the jet was measured. This was calculated over 30 periods of current and input voltage as:

$$P_{avg} = \frac{1}{30T} * \int_{t_1}^{t_2} P(t)dt,$$

where T is oscillation period, and $P(t)$ is instantaneous power in every moment t calculated as $I(t) * V(t)$ from the beginning t_1 and end t_2 of 30 periods. The measurements were performed on the electrode before the plasma jet coming out of the tube and at two gas flow rates of 1 and 2 slm. The calculated values present an

average power that the power source gives to the plasma jet (Fig. 7 left axis) and represents the ‘real’ power input into plasma. In order to link electrode voltage and power and to facilitate comparison to the other experimental data, we calculated V_{RMS} values as a function of input DC voltage (Fig. 7 right axis). The RMS values were calculated for 30 periods assessing several $V(t)$ signals at the same DC voltage in order to estimate differences. The measurements were performed on the electrode at two gas flow rates of 1 and 2 slm. It was observed that the power was not influenced by the gas flow rate but was instead dependent on the DC input voltage and provided powers in the range of 0.1–1.5 W. The power that is transferred from plasma to the treated samples is somewhat lower than calculated power since part is always lost.

Optical emission spectroscopy was used as a plasma diagnostic tool. A typical spectrum of He discharge of an APPJ at gas flow rate of 2 slm where a jet was positioned above the saline solution target is presented in Fig. 8. The spectrum was recorded in a wide range

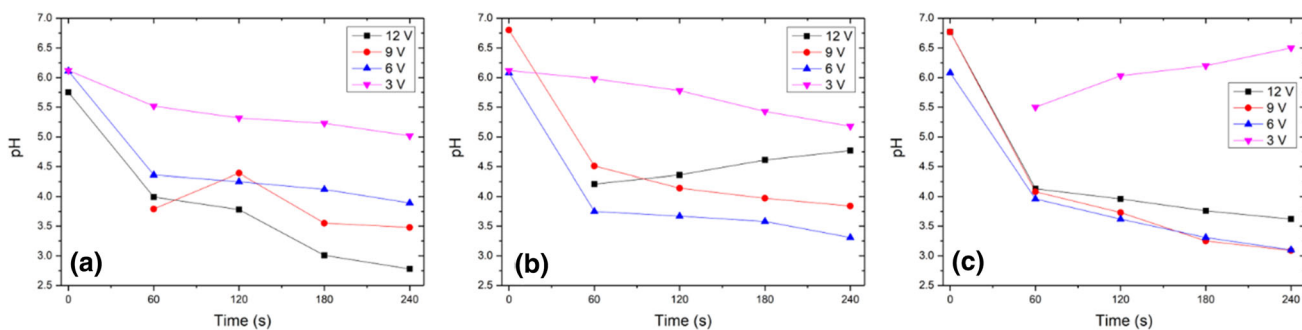


Fig. 5 pH values of He APPJ treated saline for gas flow of a 2 slm, b 1.5 slm and c 1 slm

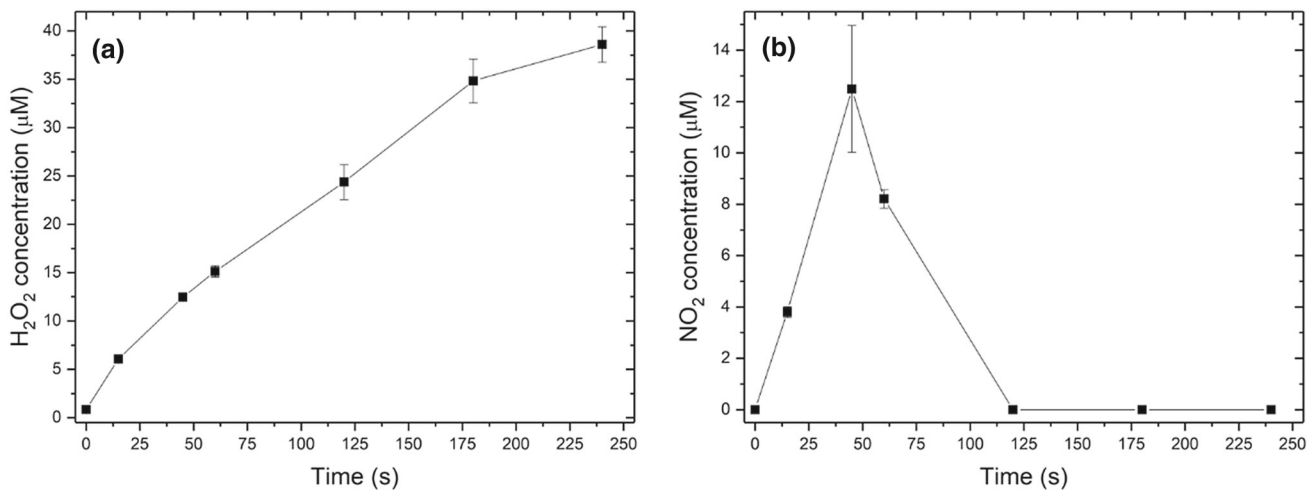


Fig. 6 Concentration of reactive species: a H₂O₂ and b NO₂⁻ with respect to treatment time

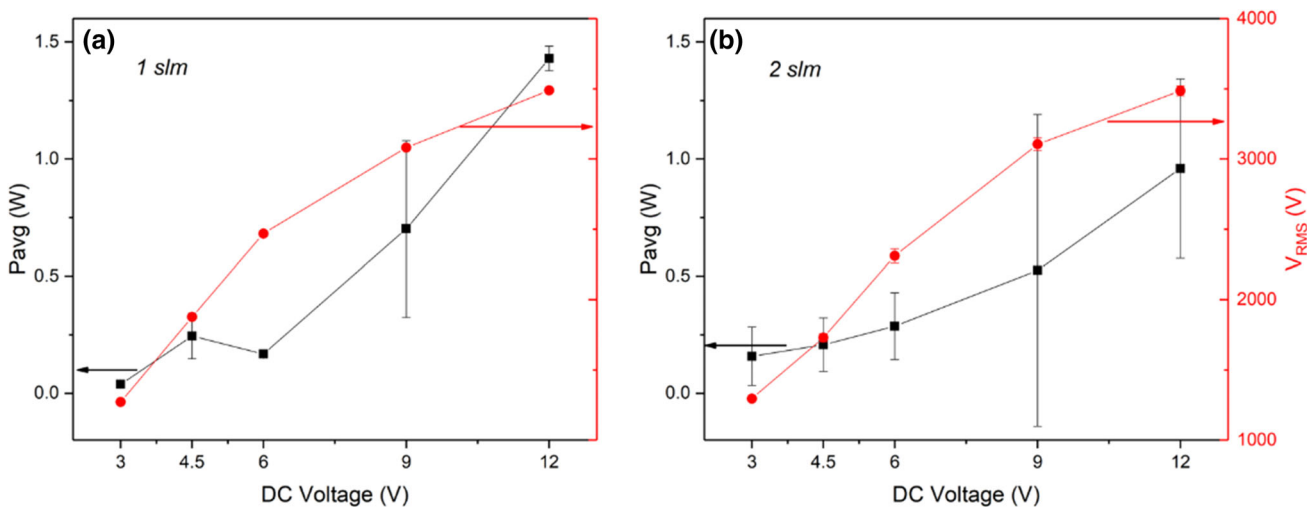
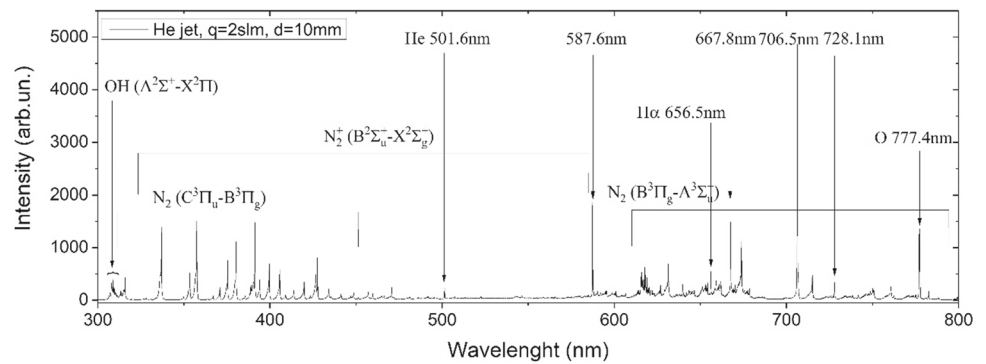


Fig. 7 Average input power to the plasma jet for He gas flows of a 1 slm and b 2 slm

Fig. 8 Optical emission spectroscopy of the He APPJ with a characteristic spectrum generated during the treatments above the liquid medium



of wavelengths, between 300 and 800 nm. The characteristic spectrum of excited species has already been assessed for this kind of plasma jet [33–35]. The most intense lines came from the molecular OH (A–X) band, atomic lines of He and O, and molecular bands of N₂; the second positive system (SPS) and the first positive system as well as from the nitrogen ion—the first negative system (FNS) [36, 37]. Excited He atoms were produced from the ground state neutrals in the working gas used in the system. At the same time, OH and N₂ bands and atomic O lines and H α line were present since the experiments were conducted in an ambient air (with some humidity) and in contact with the saline solution. Neutral species from the surrounding air were mixed with the helium flow and therefore participated in gas phase reactions induced by plasma [38, 39]. In contrast to the case where solid NaCl was treated [40], the spectrum obtained with saline solution did not show any additional lines from Na (or Cl). This suggests that these species were not excited in the gas phase above the water for the plasma source to excite them.

Additional analysis regarding line intensity was performed on specific atomic and molecular lines for different DC input voltages used in the experiment (3, 4.5, 6, 9 and 12 V) at two gas flows (1 and 2 slm). The intensities at different discharge parameters with two emission lines from N₂ SPS (337.1 nm and 315.9 nm), head line from FNS N₂⁺ (391.3 nm), the strongest molecular OH line (309 nm), H α (656.6 nm) and O atom line (777.4 nm) are presented in Fig. 8, while He line (706.5 nm) intensities are depicted in Fig. 9. All line intensities are normalised to the same recording conditions and corrected for spectral efficiency of the system, thus allowing direct intensity comparison between different lines. The position of the jet and the distance to the bottom of the 96-well plate were the same as for the treatments of bacteria. There was an increase in intensities for all observed lines when the source power (DC voltage) was increased. The increase of He flow had a minor influence on line intensities, resulting in a somewhat higher line intensity. In all cases, there was a stronger or weaker ‘jump’ between the emission intensities recorded for 6 V and 9 V. This change in peak values occurred due to the change in plasma regime since, as observed with the naked eye, the plasma channel did not connect to the surface of the saline until the 9 V

were reached [41]. Therefore, the intensities recorded for voltages below 9 V can be regarded as free-standing jet cases, while for the voltages of 9 V and 12 V, plasma plume was in contact with the liquid surface.

The highest line intensities belong to the mainline of N₂ SPS, and these intensities have pronounced increments between 6 and 9 V input voltage (Fig. 9a and b). The second strongest line of the same band has a much lower increase in intensity. However, excitation of both of the excited levels in N₂ probably happened through electron collisions with the ground state or excited N₂ molecules [35, 42]. The increasing line intensity tendency is in accordance with the dependence observed with similar jet configurations [34]. The intensity of the strongest of FNS N₂⁺ lines at 391.3 nm also increased with DC voltage, yet much less than the 337.1 nm line (Fig. 9a and b). This line comes from the excited state of N₂⁺ ions that were efficiently produced in the Penning ionisation process, involving He metastables [43] and the direct electron impact ionisation process [42]. Consequently, an increase in He flow made the emission of the 391.3 nm line rise. On the other hand, lines from the OH band and H α came from dissociation of water vapour molecules in plasma [35, 44]. In this jet configuration, the amount of water vapour present in the surrounding air was sufficient to produce several excited species of OH and H visible in the emission spectrum. An increase in the He flow and discharge voltage resulted in the increase of OH emission intensity (Fig. 9c and d), which has been observed before [34, 45]. The atomic O (777.4 nm) line exhibited similar behaviour. Production of both OH and O species is important when it comes to the treatment of bacteria.

The intensity trend of the He line at 706.5 nm was similar to that of other spectral lines and is presented in Fig. 10. As expected, line intensity was observed to increase when we increase either working gas flow or DC input voltage. This He line is the most intense compared to other lines observed in the spectrum (Fig. 10). The result is due to amount of He and its mixture with air present in the plasma plume.

Observing all line intensities analyzed here, one can conclude that within the range of voltages varied in the experiment, there is a steady increase of the line intensity with increase in applied voltage. This reflects a fact

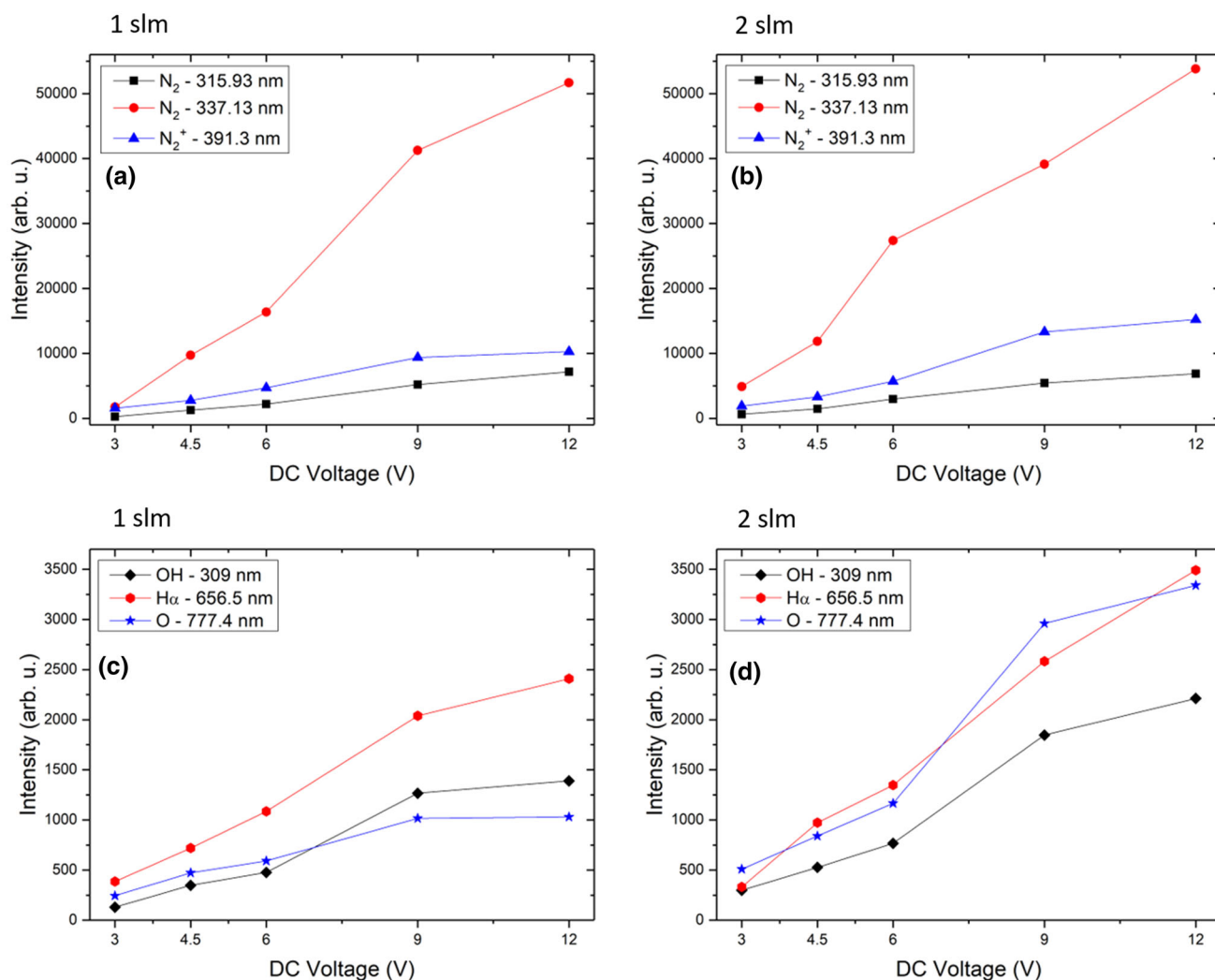


Fig. 9 Optical emission spectroscopy: intensities of certain atomic and molecular lines of nitrogen (**a** for 1 slm and **b** for 2 slm), and hydrogen, oxygen and hydroxyl **c** for 1 slm and **d** for 2 slm)

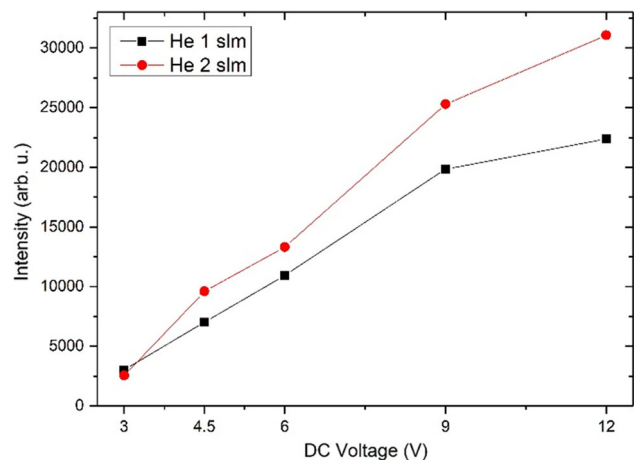
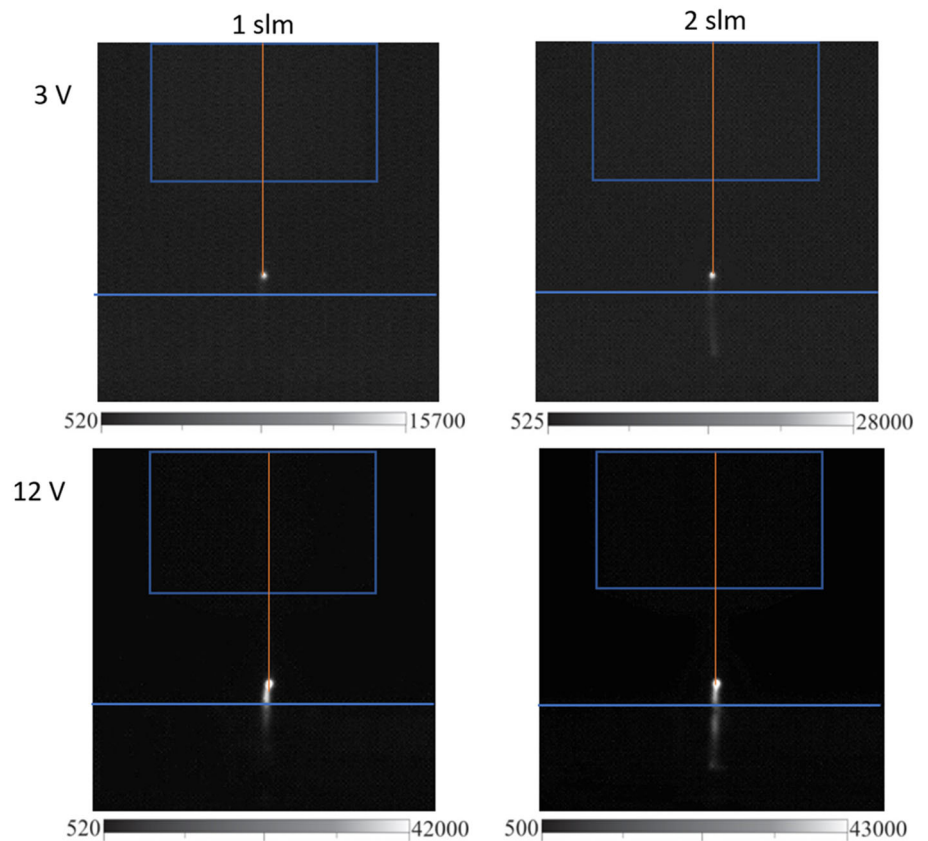


Fig. 10 He 706.5 nm line intensities for different gas flow and voltages

that for all lines, i.e. processes related to specific emission, concentration of excited species continuously rises with voltage increment, without any abrupt changes. Hence, we can say that both voltage and flow changes applied here do not influence plasma chemistry but only concentration of species involved in the processes.

Discharge imaging can provide information about the way plasma plume forms and how it propagates between electrode and bacteria-containing medium. Additionally, a relative abundance of active species can be roughly assessed through emission intensity as brighter intensity corresponds to more emitting particles. For this, ICCD imaging was employed with time-averaged images of the streamer structure obtained for all He flows and DC input powers. Typical results are presented in Fig. 11 for 1 slm and 2 slm, at only the lowest (3 V) and highest (12 V) DC voltages used in the experiments. Similar to optical emission measurements, the jet position and its distance to the liquid surface in

Fig. 11 Images of the discharge structure at different flows and DC voltages from the power supply. A thin vertical line represents a pin electrode protruding from the body of the jet (rectangle shape). The horizontal line in the lower part of the images signifies the upper edge of the 96-well plate



432 the 96-well plate were the same as for bacteria treat-
 433 ments. For all conditions, a ball-shaped bright plasma
 434 was visible on the tip of the pin electrode with a plasma
 435 plume extending towards the target. For the lowest DC
 436 voltage employed, a weak plasma channel existed only
 437 for the 2 slm flow of He. Obviously, for $V_{DC} = 3\text{ V}$ and
 438 1 slm of He, the field attained at the electrode with these
 439 power supply conditions was not enough to achieve suf-
 440 ficient ionisation in the whole volume between the jet
 441 and sample surface. At the highest power, i.e. DC volt-
 442 age of 12 V, a streamer-like plasma channel with strong
 443 emission bridged the distance from the electrode tip to
 444 the liquid surface. After processing all images recorded
 445 by subtracting the background intensity level, it was
 446 determined that in all applied conditions, except for 3 V
 447 at 1 slm, the plasma plume reached the liquid surface,
 448 meaning that streamer length in this range of condi-
 449 tions did not depend on either voltage or helium flow.
 450 This indicates that the medium chemistry of bacteria
 451 deactivation depends on streamer forming behaviour,
 452 which almost doubles the procedure's efficacy.

453 4 Conclusion

454 To optimise bacteria deactivation in media and obtain
 455 sterilisation with plasmas, a parameter study involv-
 456 ing a large number of experiments using different

457 plasma conditions was performed. It included moni-
 458 toring the viability of different bacteria strains with
 459 respect to several plasma diagnostics measurements.
 460 This research clearly shows non-thermal helium APPJs'
 461 ability to deactivate four standard strains of bacteria
 462 used in such experiments. The deactivation effects of
 463 the plasma jet were significant and dependant on the
 464 bacterial strain, exposure time and plasma configura-
 465 tion (gas flow rate and input DC power unit voltage).
 466 The obtained results are expected and indicate that *E.*
 467 *coli* is deactivated faster than other strains. Generally,
 468 all bacterial strains—*E. coli*, *S. aureus*, *B. stearother-*
 469 *mophilus* and *B. subtilis*—follow the same deactivation
 470 trends. The only discrepancy is in the optimal param-
 471 eters for deactivation of *B. stearothermophilus*, where
 472 optimal deactivation is reached at lower flow rate levels.
 473 This might be because of the bacterial strain's proper-
 474 ties and its response to the changing environment by
 475 interacting plasma. The interaction of plasma and bac-
 476 teria suspension (saline) was twofold—it changed the
 477 concentration of reactive species and pH in the solu-
 478 tion with bacteria. This RONS species (He, O, N, H,
 479 photons) generated in the gas phase and high-energy
 480 electrons and ions were interacting with the liquid. As
 481 a result of the combined action of produced reactive
 482 species and chemical reactions, which also influenced
 483 the pH in the liquid phase, increased bacteria deacti-
 484 vation efficacy. Combining all the chemically initiated
 485 processes managed to sterilise given bacterial strains in
 486 a medium in fairly short treatment times, maximum

487 efficacy was observed at high flow rates and DC input
 488 powers. More power transferred into the plasma short-
 489 ens the deactivation process. Increasing the flow rate
 490 from 0.5 to 2 slm also shortens the inactivation process
 491 as reactive species concentration in the gas phase rises.

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501 Author contributions

502 UC and ZLP conceived and planned the experiments. NH,
 503 MM, DV and MĐ performed plasma treatment of bacteria
 504 and liquid chemistry analyses along with the interpretation
 505 of those results. NS, KS and AJ performed plasma diag-
 506 nostics and electrical characterization along with interpre-
 507 tation of those results. AJ wrote the original draft, and all
 508 co-authors helped with manuscript revision.

509 Declarations

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